

What to do with a billion years of evolution

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PI: Joseph Ryan

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DEEPC

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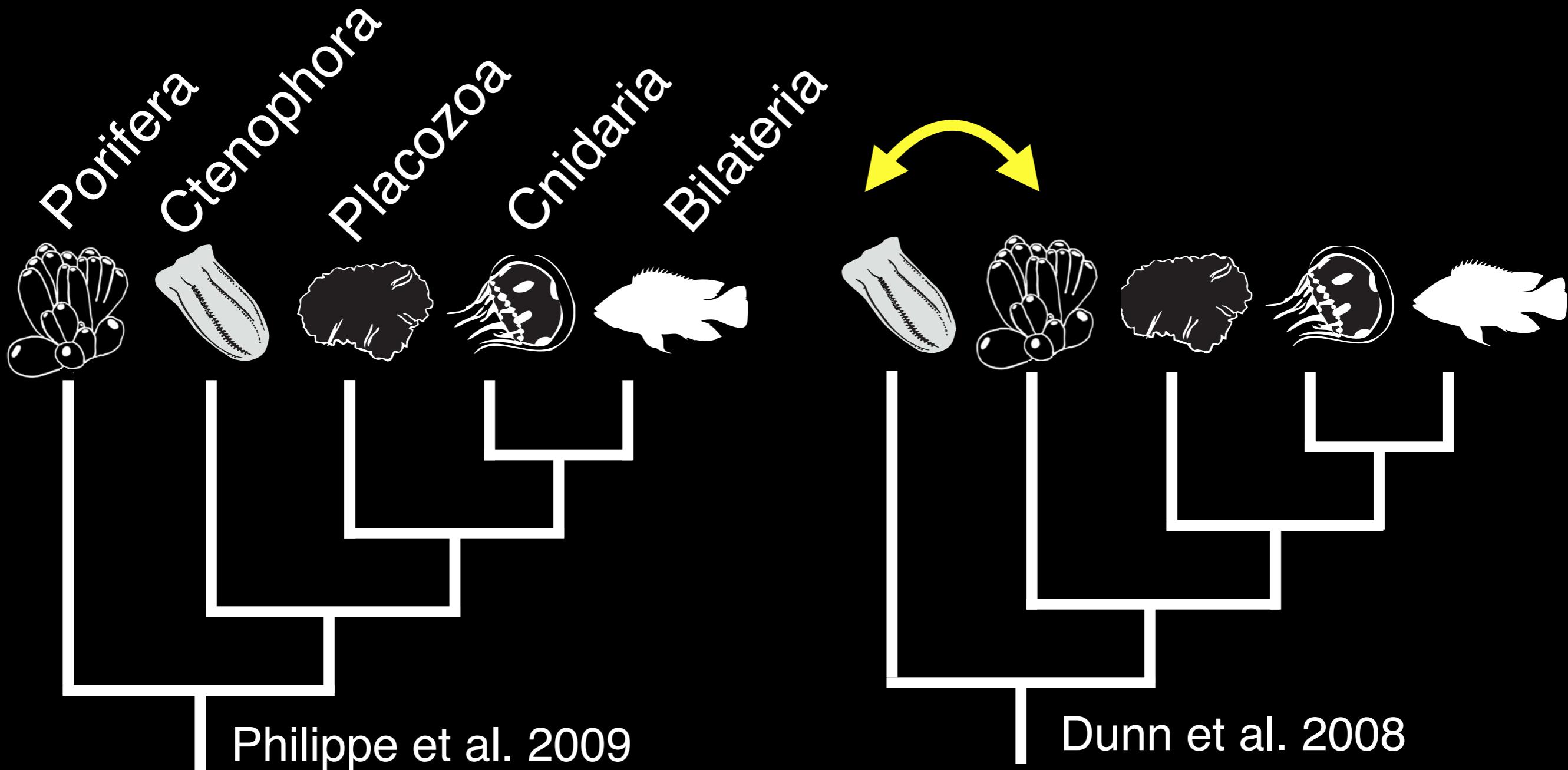
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Telissa Wilson

Warren Francis • University of Southern Denmark

Funding

NSF Dimensions of Biodiversity (1542679, 1542673, 1542597)



Pick et al. 2010

Nosenko et al. 2013

Pisani et al. 2015

Simion et al. 2017

Feuda et al. 2017

Philippe et al. 2009

Dunn et al. 2008

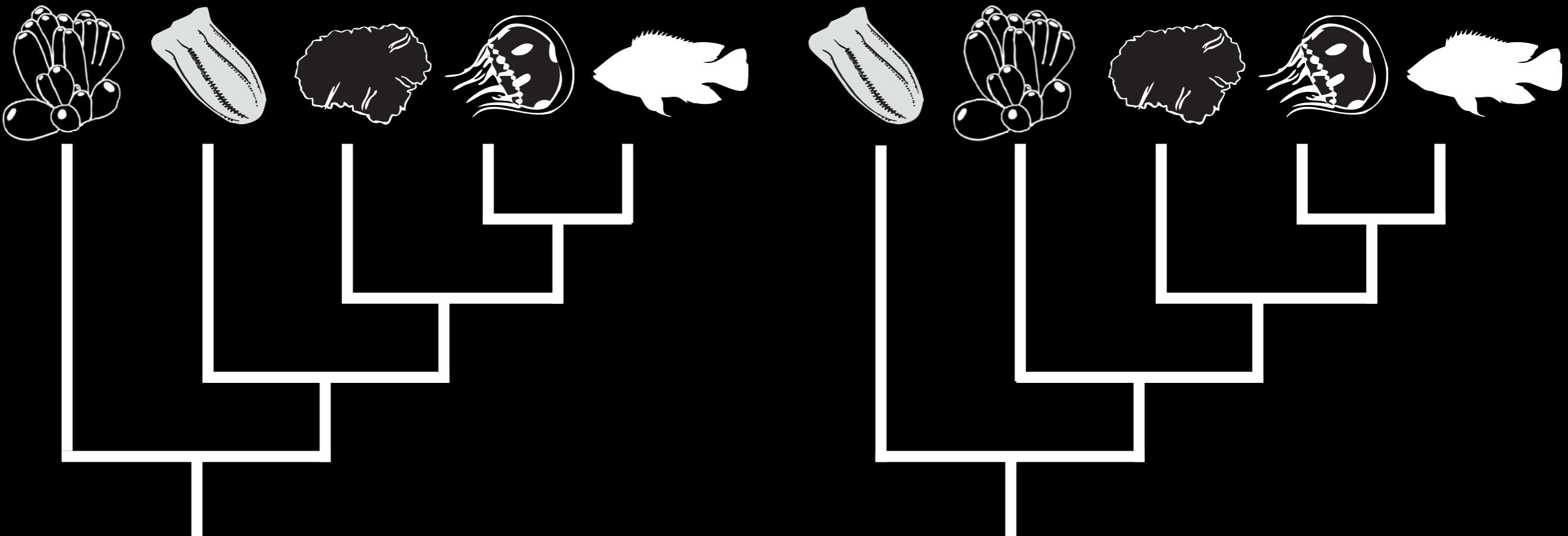
Ryan et al. 2013, Hejnol et al. 2009

Moroz et al. 2014, Chang et al. 2015

Whelan et al. 2015, Torruella et al. 2015

Borowiec et al. 2015, Arcila et al. 2017

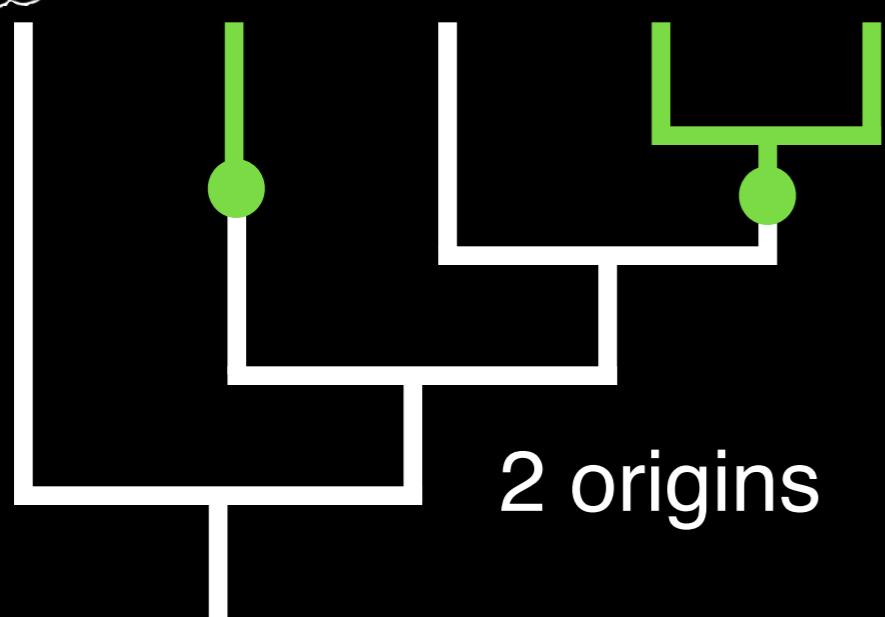
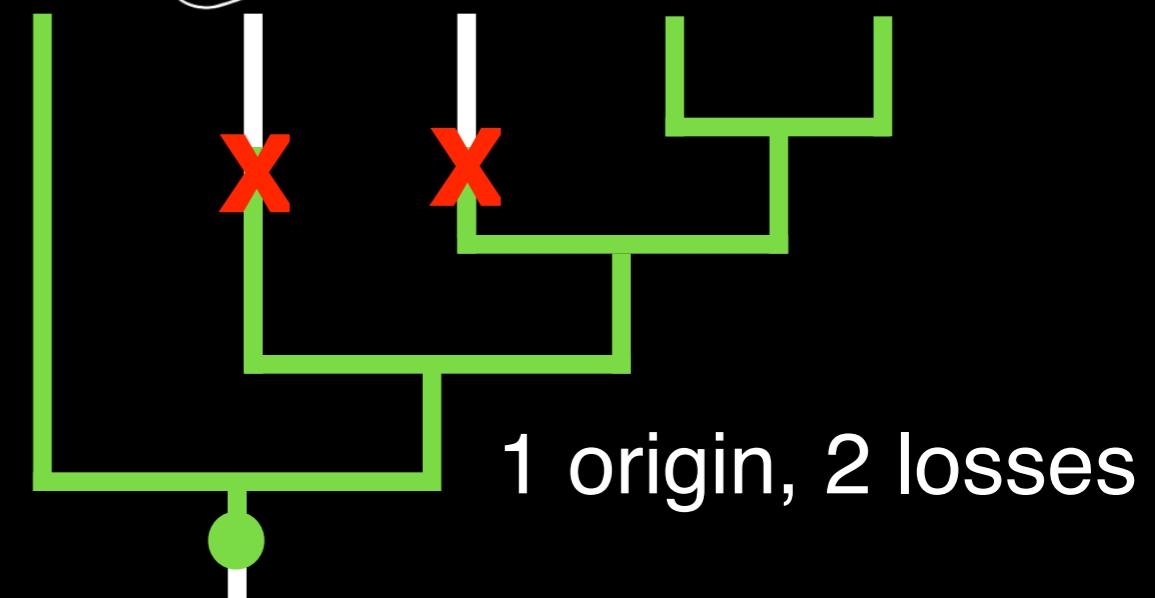
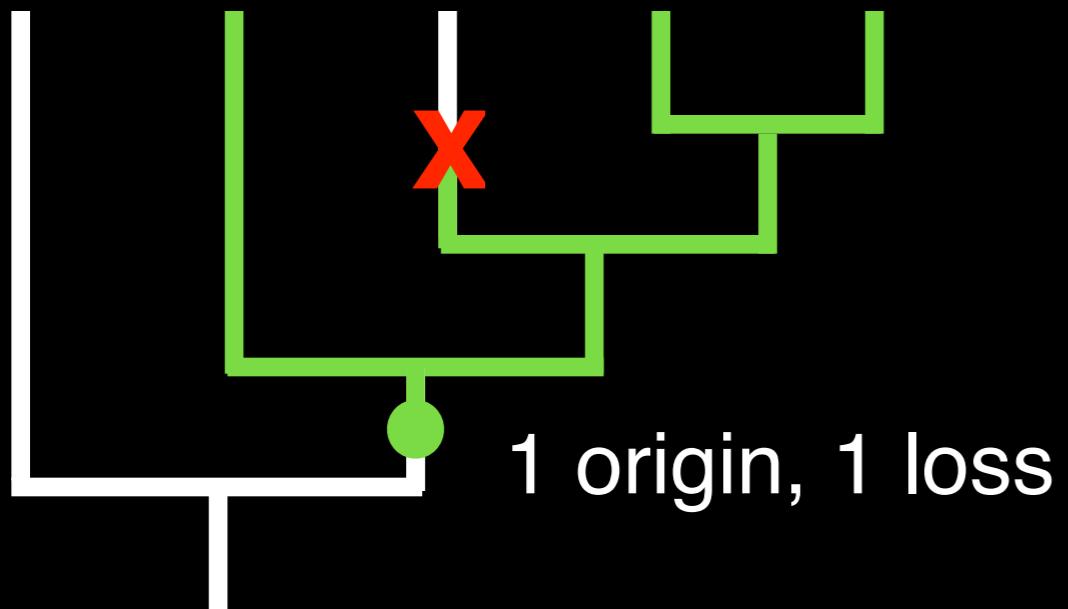
Whelan et al. 2017, Shen et al. 2017



Who cares?

Accurate species trees are important tools
for understanding evolutionary history

Evolution of neural cell types



Evolution of neural cell types

Species trees estimated over different time scales

species complex in cardinals

10 mya



shallow

introgression

rapid radiation

RADseq/UCEs

nucleotide alignments

gene tree / species tree methods

relationships among insects

500 mya



origin of multicellularity

>1 billion mya

deep

ancient splits

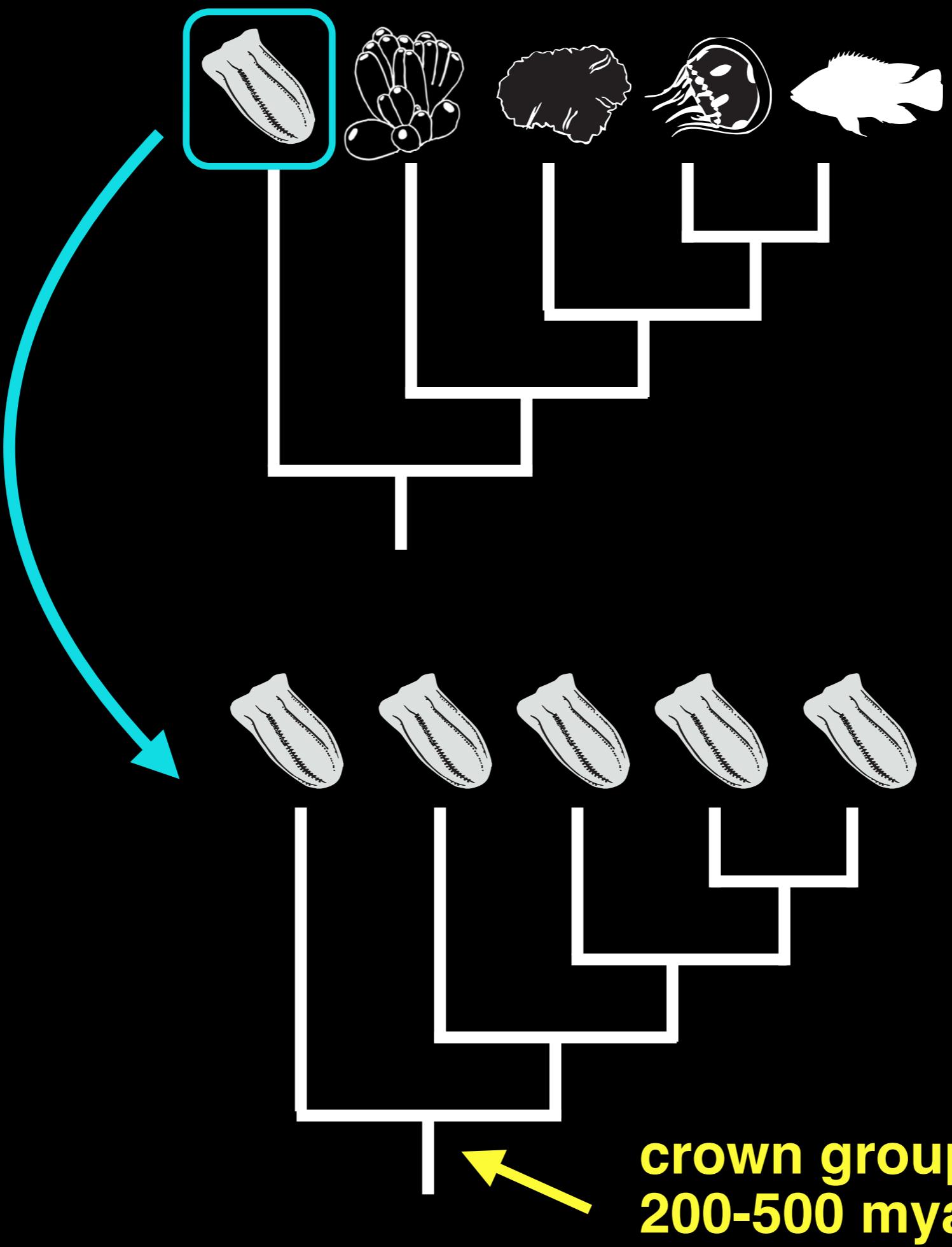
saturation

compositional heterogeneity

methods???

past time

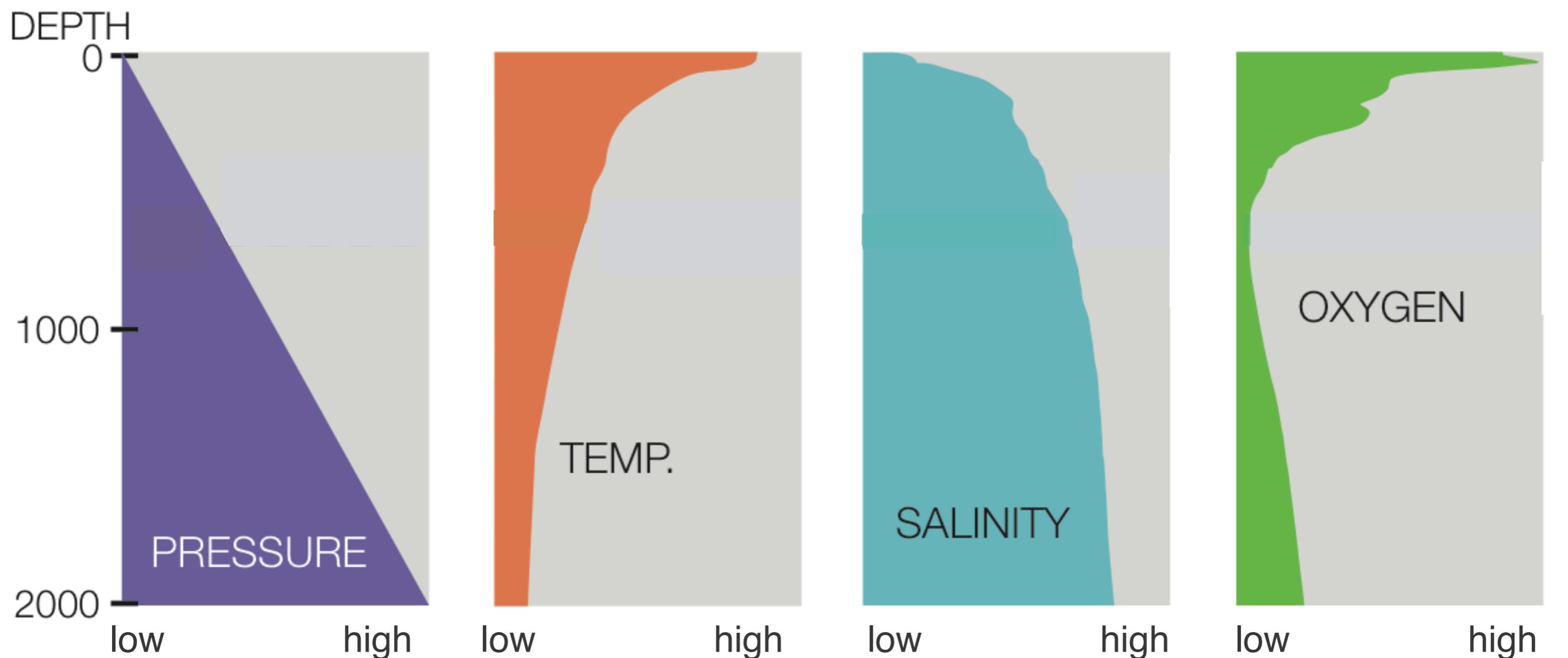
..... present





Dimensions of Biodiversity

DEEPC:
Diversity, Ecology, EcoPhysiology of Ctenophores





Dimensions of Biodiversity

DEEPC: Diversity, Ecology, EcoPhysiology of Ctenophores

Comprehensive ctenophore species phylogeny

Reconstruct ancestral depths

Identify evolutionary depth transitions

Test loci for convergent evolution to depth

Link genotype to phenotype



PHYLOTTOCOL

Promoting transparency and
overcoming bias in phylogenetics

OUTLINE
DECISIONS
BEFORE
RUNNING
ANALYSES

TRANSPARENCY
ACCOUNTABILITY
reproducibility
collaboration
education

Joseph Ryan & Melissa DeBiasse

<https://peerj.com/preprints/26585/>

<https://github.com/josephryan/phylotocol>



#1542597

Data processing & analysis pipeline

RNA-Seq



Data processing & analysis pipeline

RNA-Seq

Assemble & translate transcriptome



CCGGAACGAGCACAGCCA



PGRAQP

Data processing & analysis pipeline

RNA-Seq

Assemble & translate transcriptome

Filter non-target sequences

```
>Trinity_1234  
PGRAQPPGRAQPPGRAQP  
PVRNQADCHAQIPGMAFTI  
PGRAQPPGRAQPPGRAQP  
PVRNQADCHAQIPGMAFTI
```



BLAST database

metazoa

non-metazoa

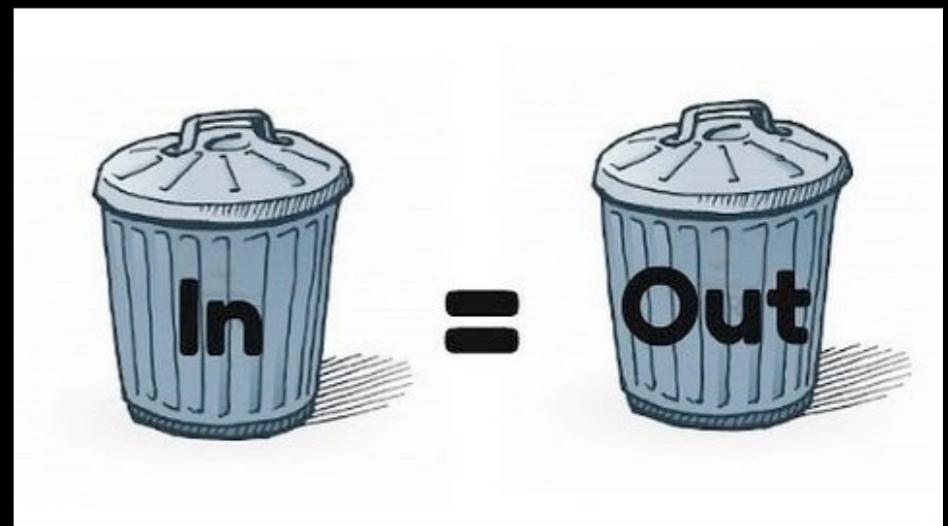
Data processing & analysis pipeline

RNA-Seq

Assemble & translate transcriptome

Filter non-target sequences

Identify orthologous loci



Orthologs: evolved from a common ancestor by speciation



Orthologs: evolved from a common ancestor by speciation

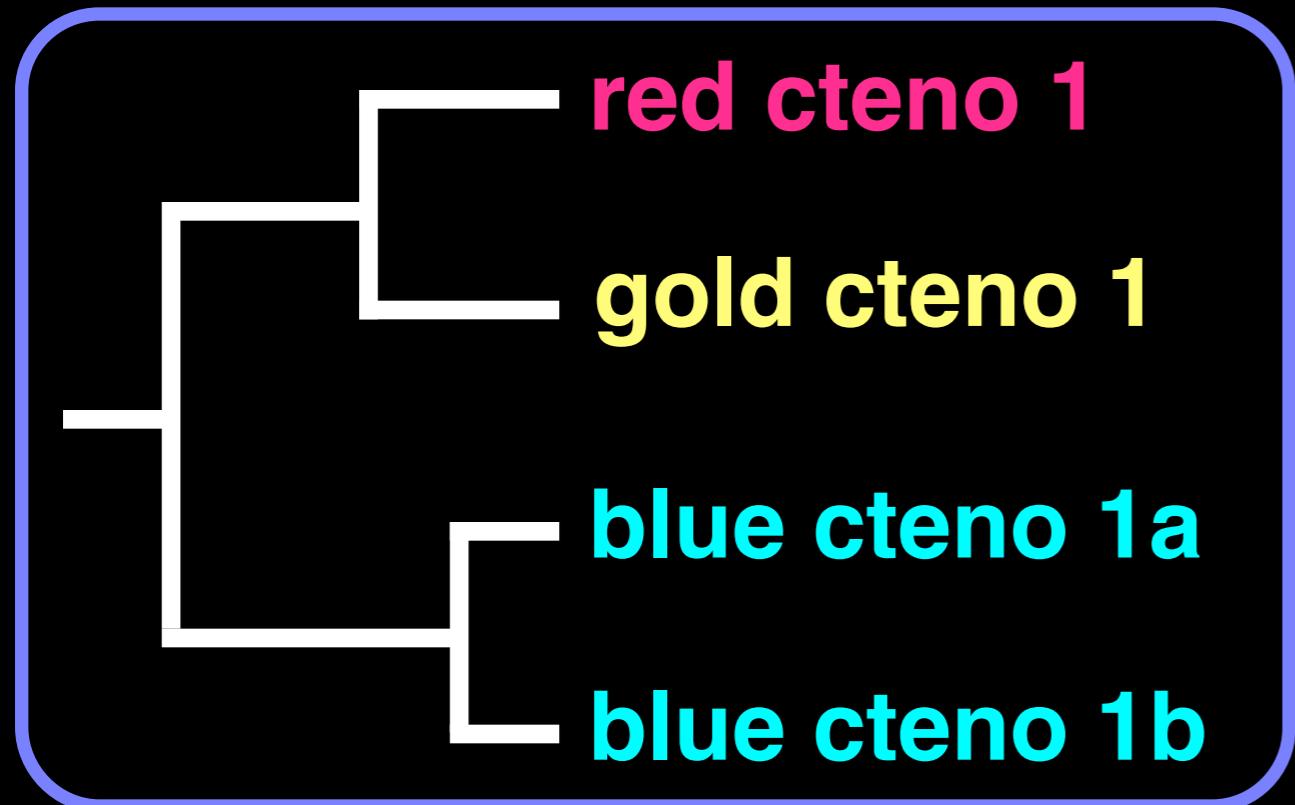
Paralogs: produced through a duplication event



Orthologs: evolved from a common ancestor by speciation

Paralogs: produced through a duplication event

Orthogroup: orthologs and paralogs from MRCA



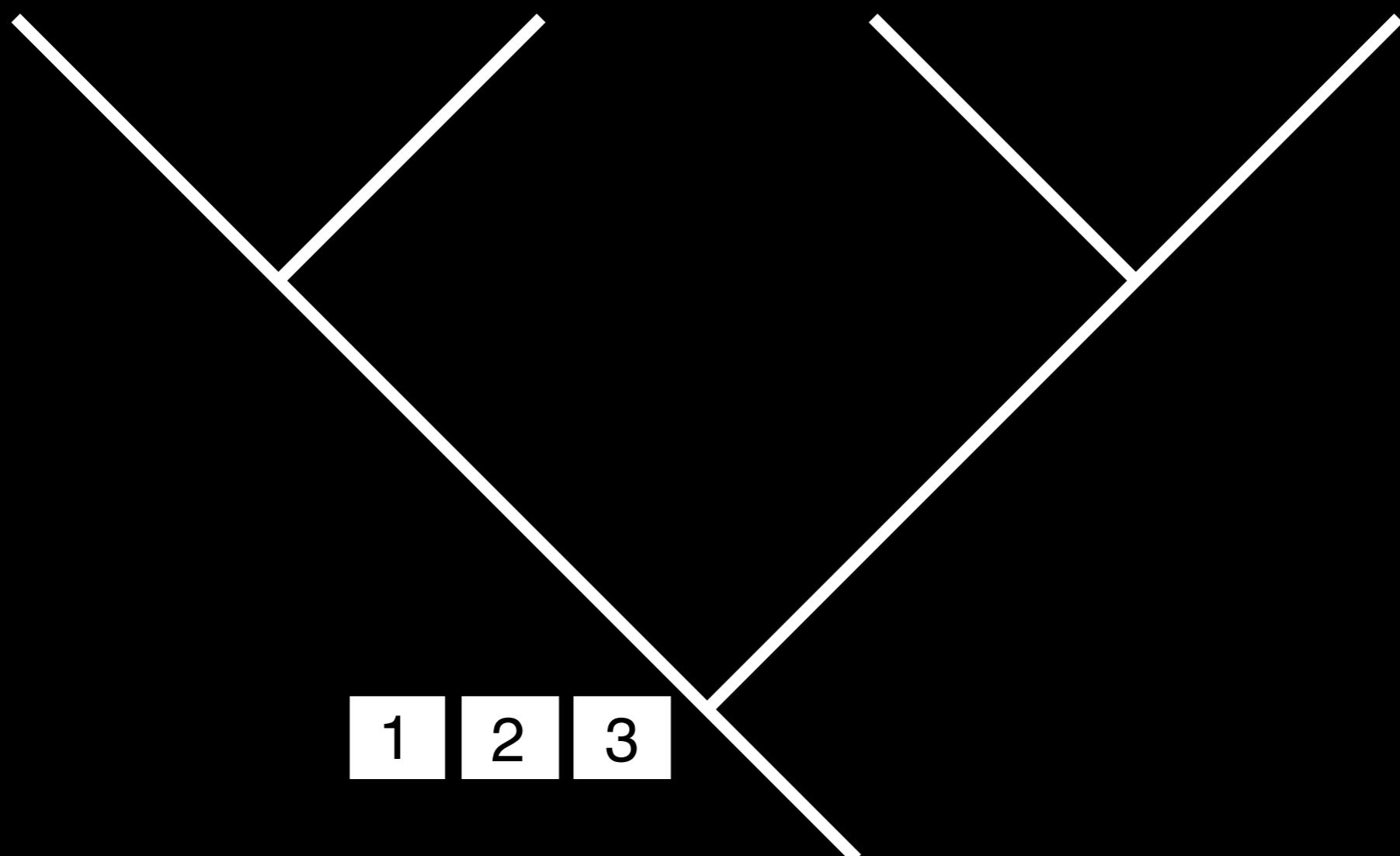
Orthogroup

1	2	3
---	---	---

1	2	2
---	---	---

1	3
---	---

1	3	3
---	---	---



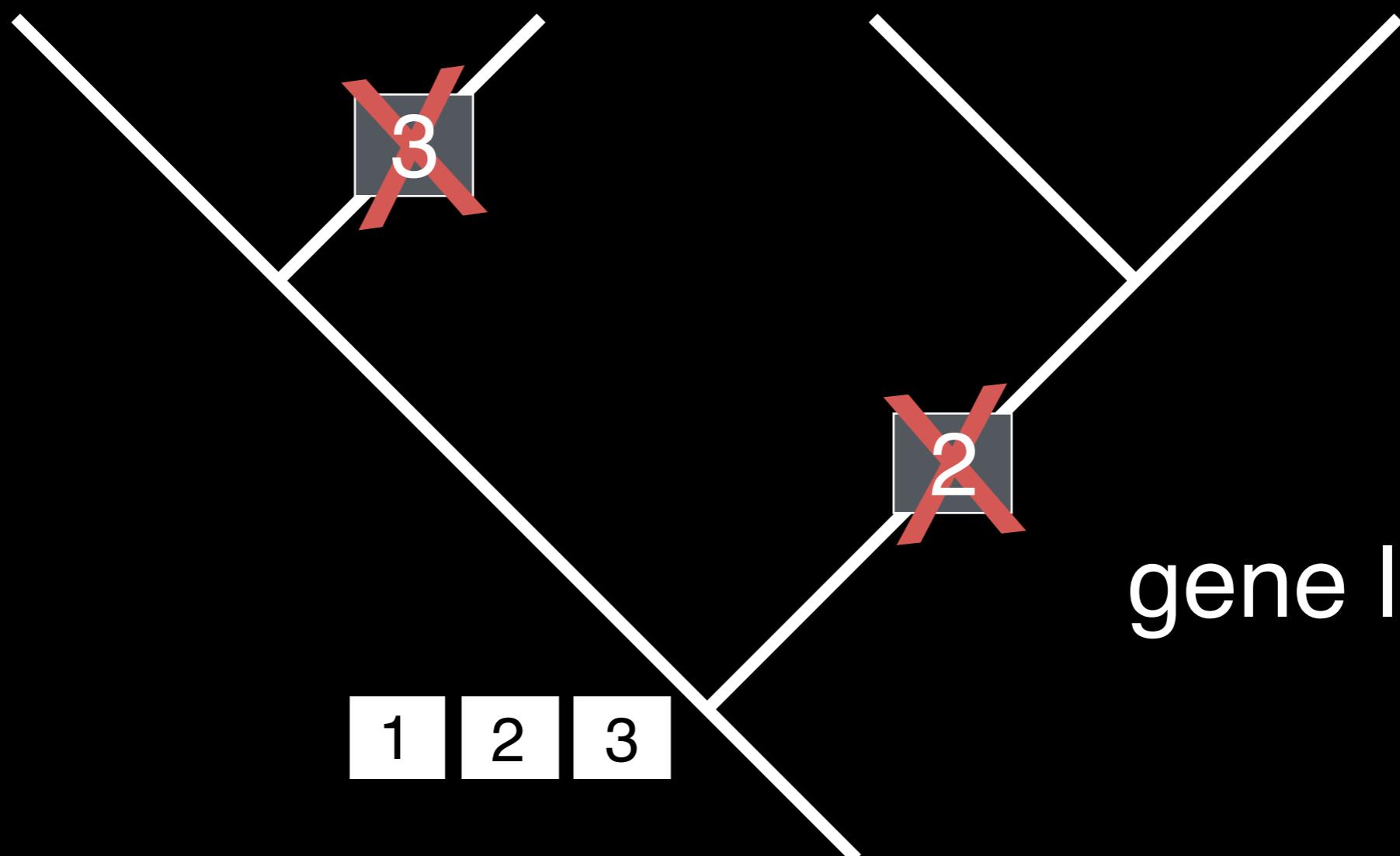
colors = species
numbers = genes

1	2	3
---	---	---

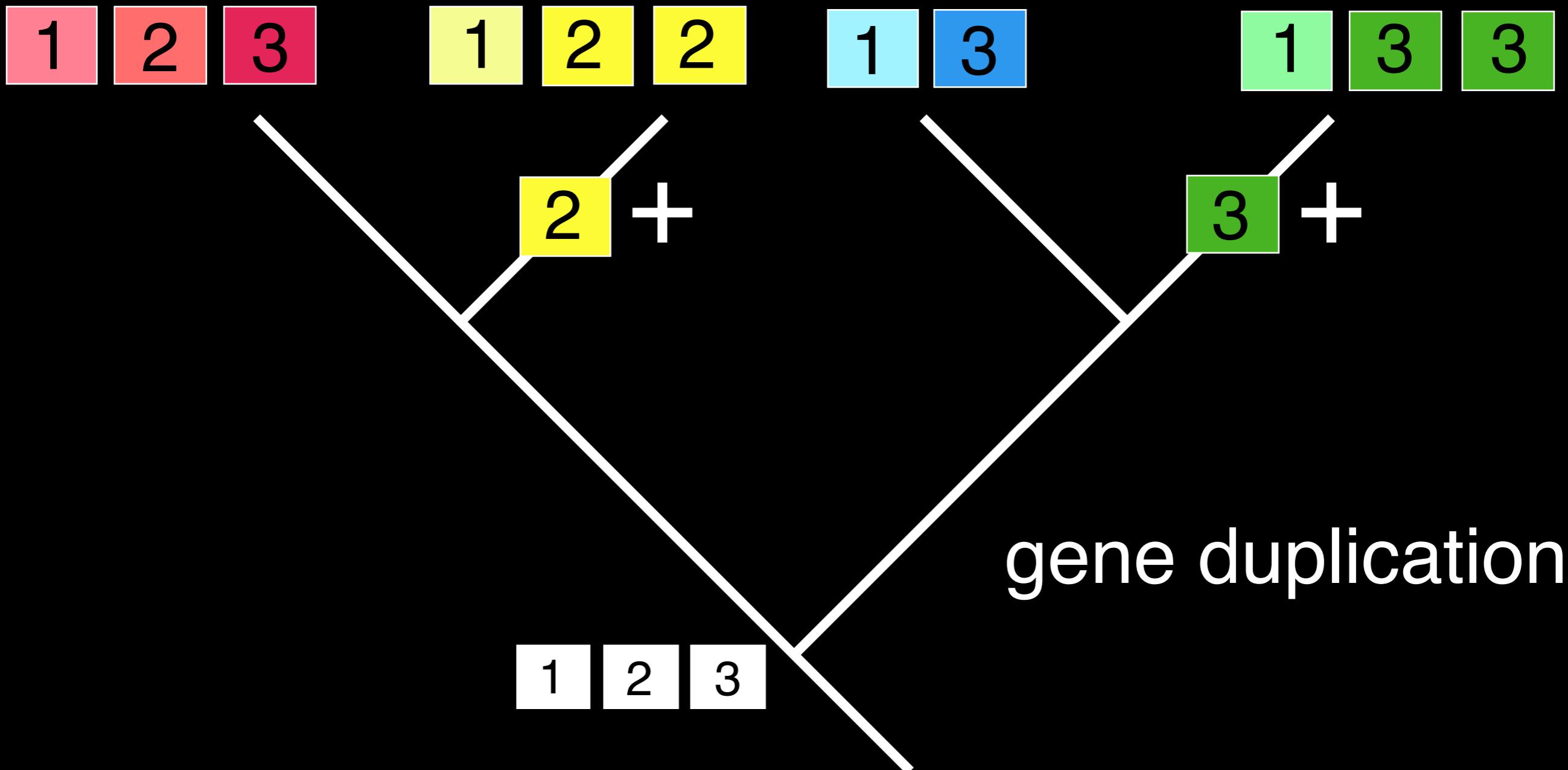
1	2	2
---	---	---

1	3
---	---

1	3	3
---	---	---



colors = species
numbers = genes



gene duplication

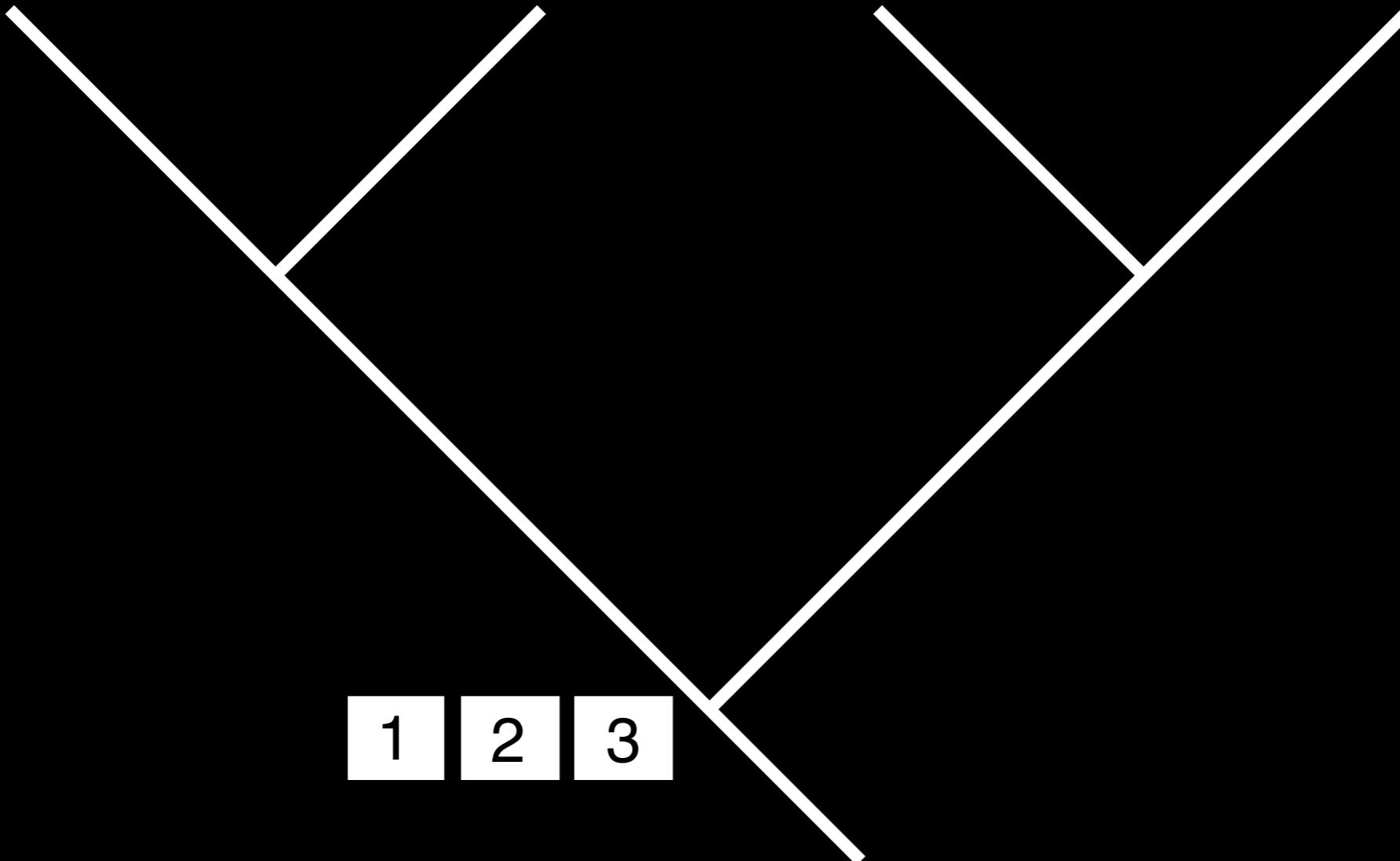
colors = species
numbers = genes

1	2	3
---	---	---

1	2	2
---	---	---

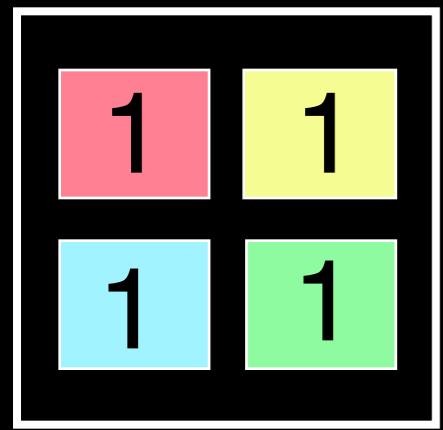
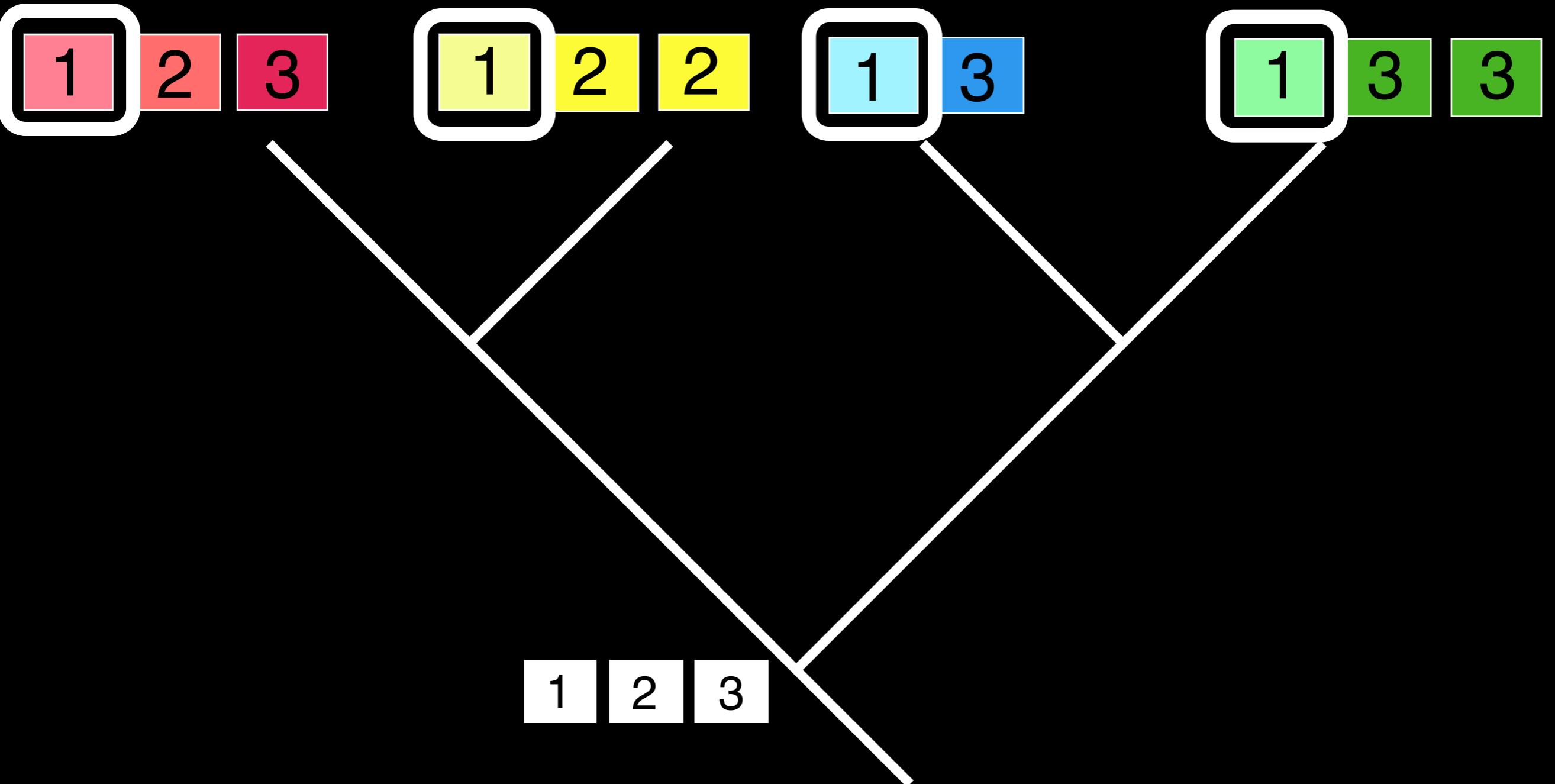
1	3
---	---

1	3	3
---	---	---

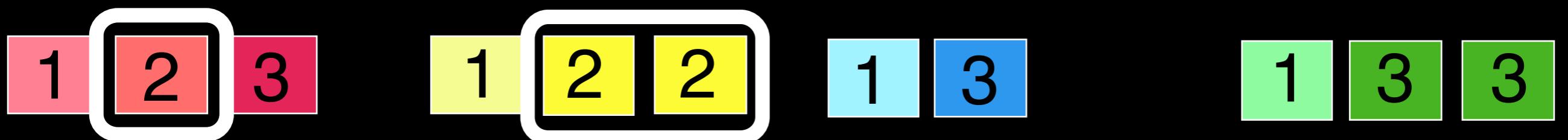


1	2	3
---	---	---

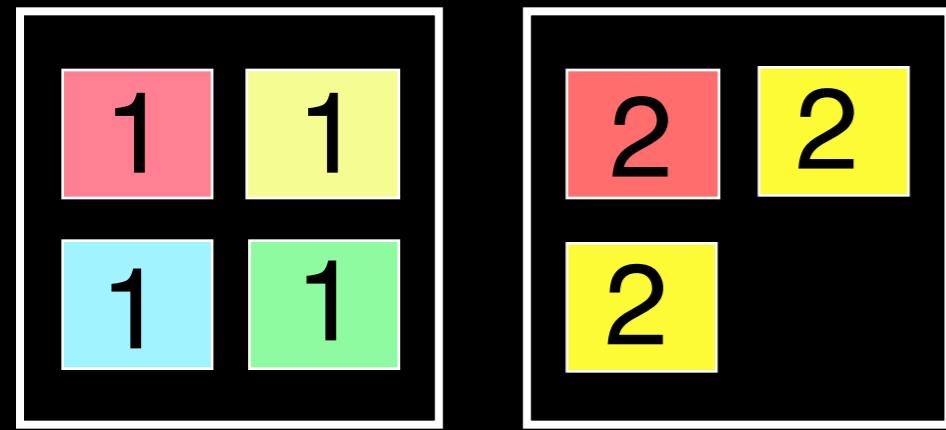
what are the ideal orthogroups?



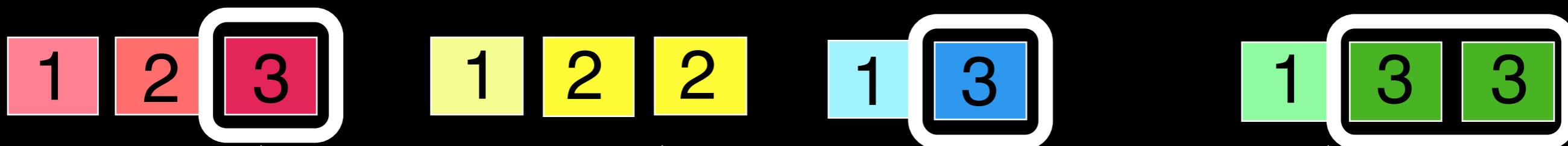
orthogroup 1



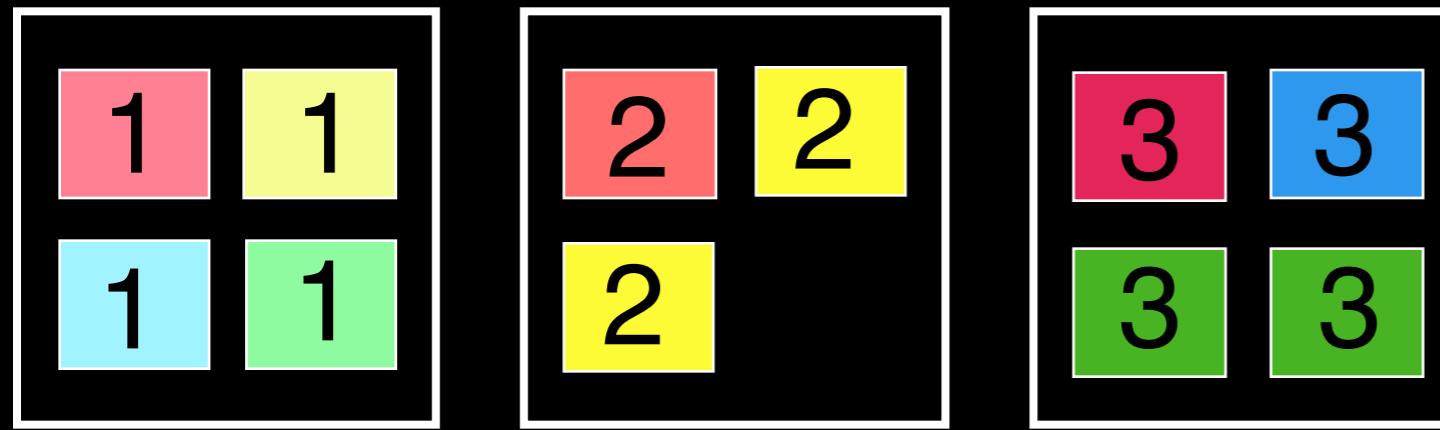
1 2 3



orthogroup 2



1 2 3



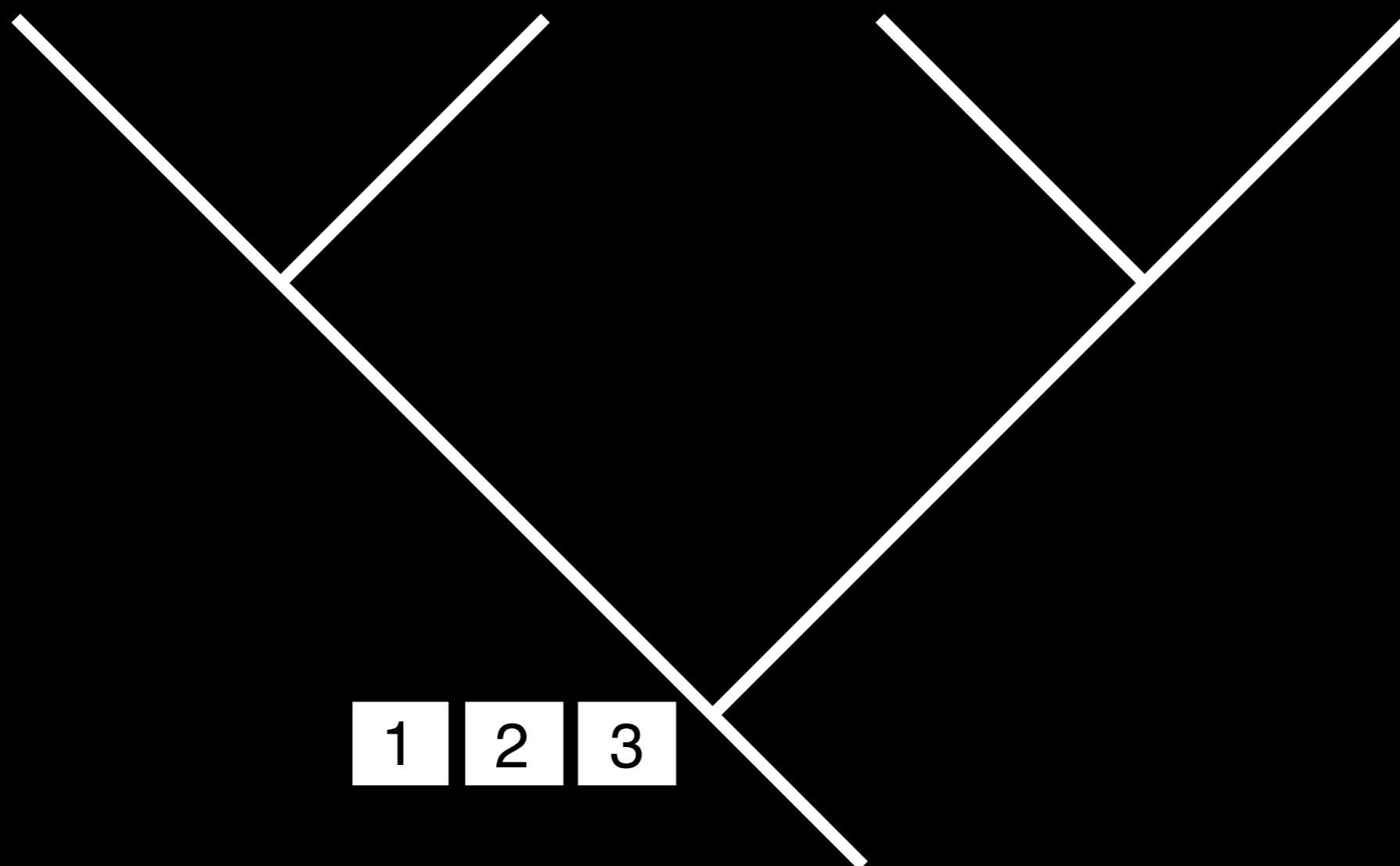
orthogroup 3

1	2	3
---	---	---

1	2	2
---	---	---

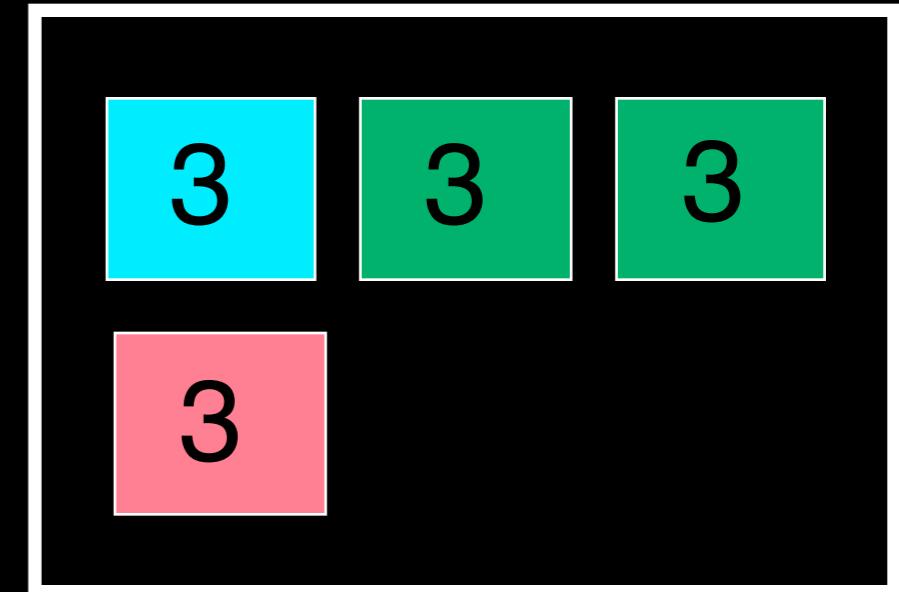
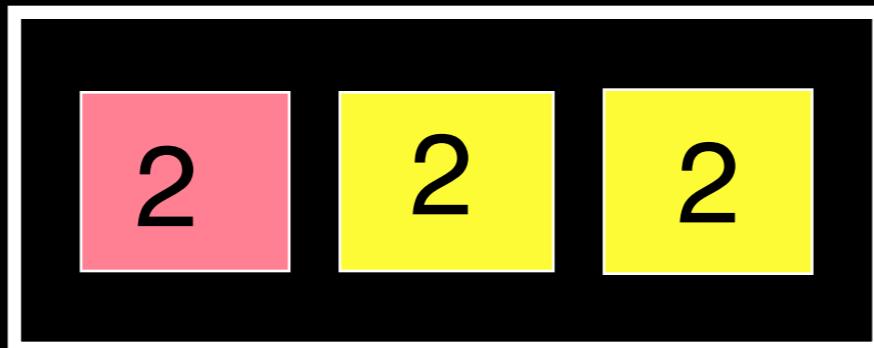
1	3
---	---

1	3	3
---	---	---



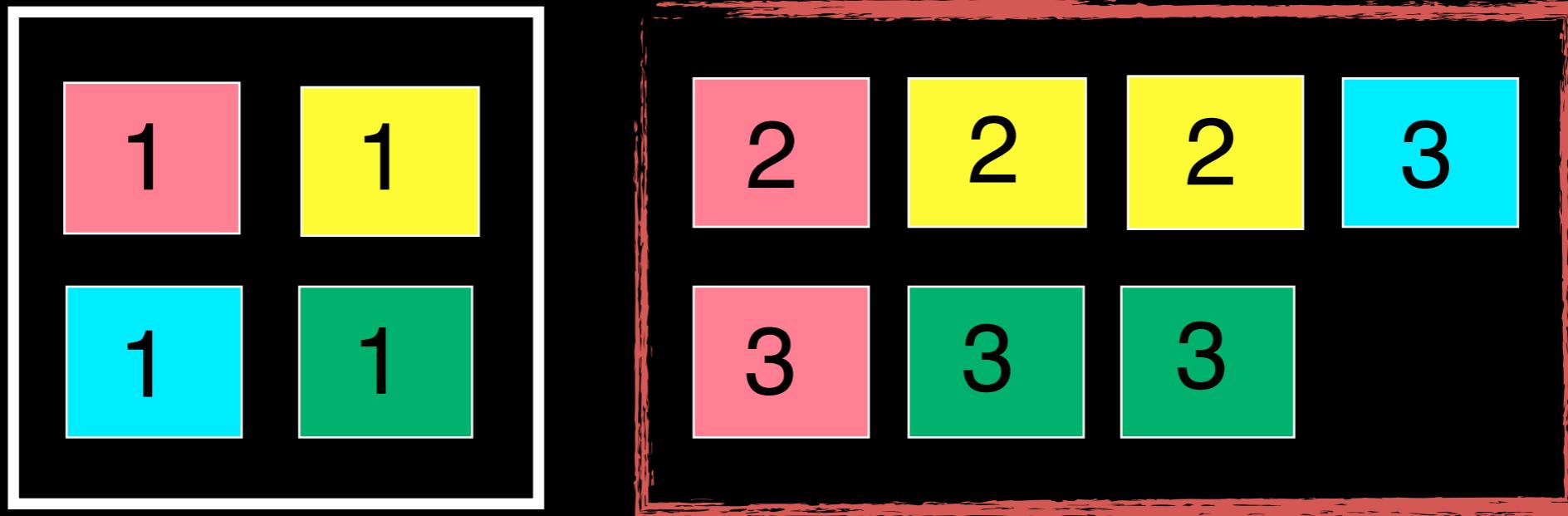
identifying orthology complicated by gene dup. &
loss, rate variation, HGT, contamination, etc

Splitting



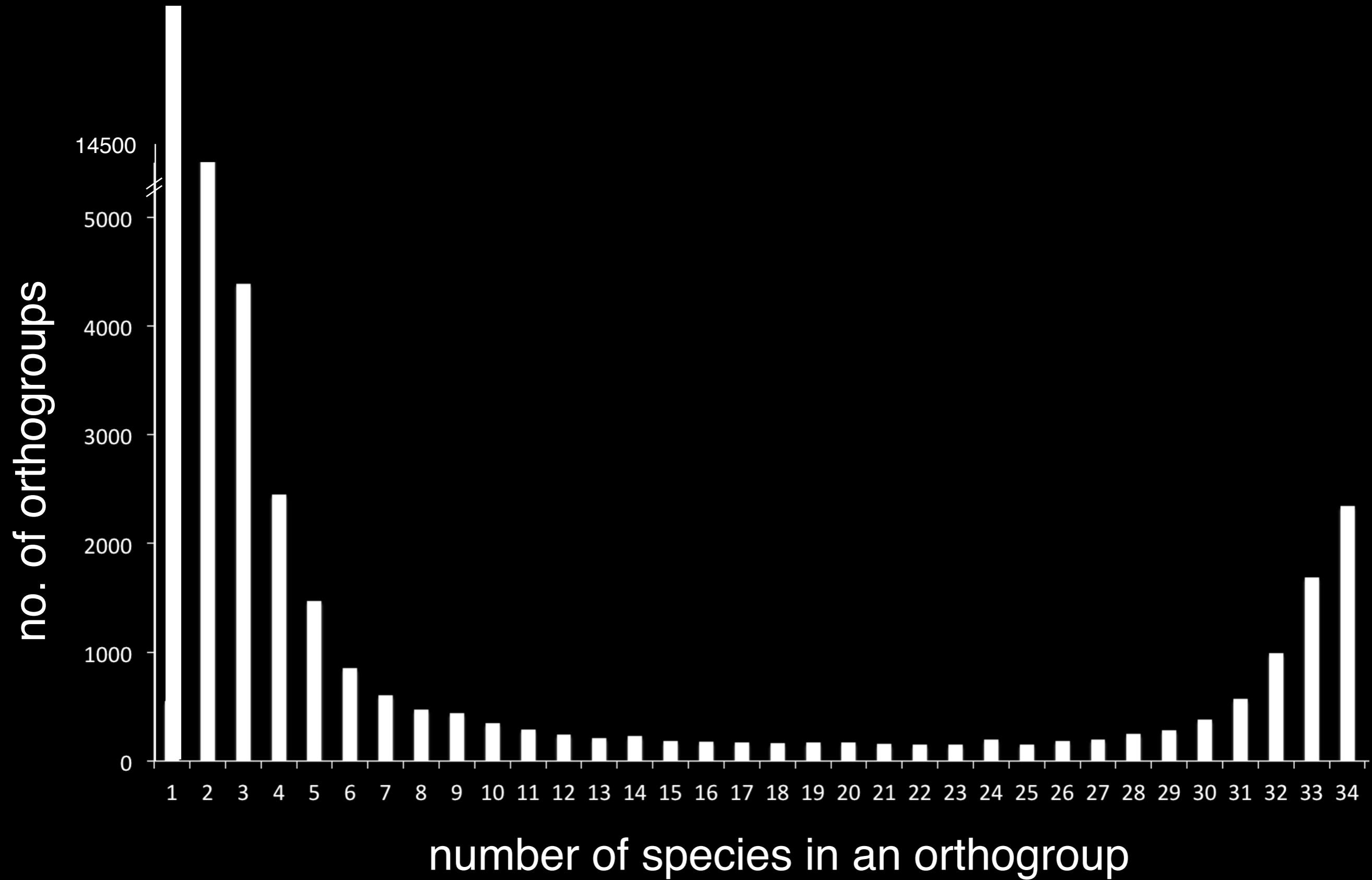
colors = species
numbers = genes

Lumping

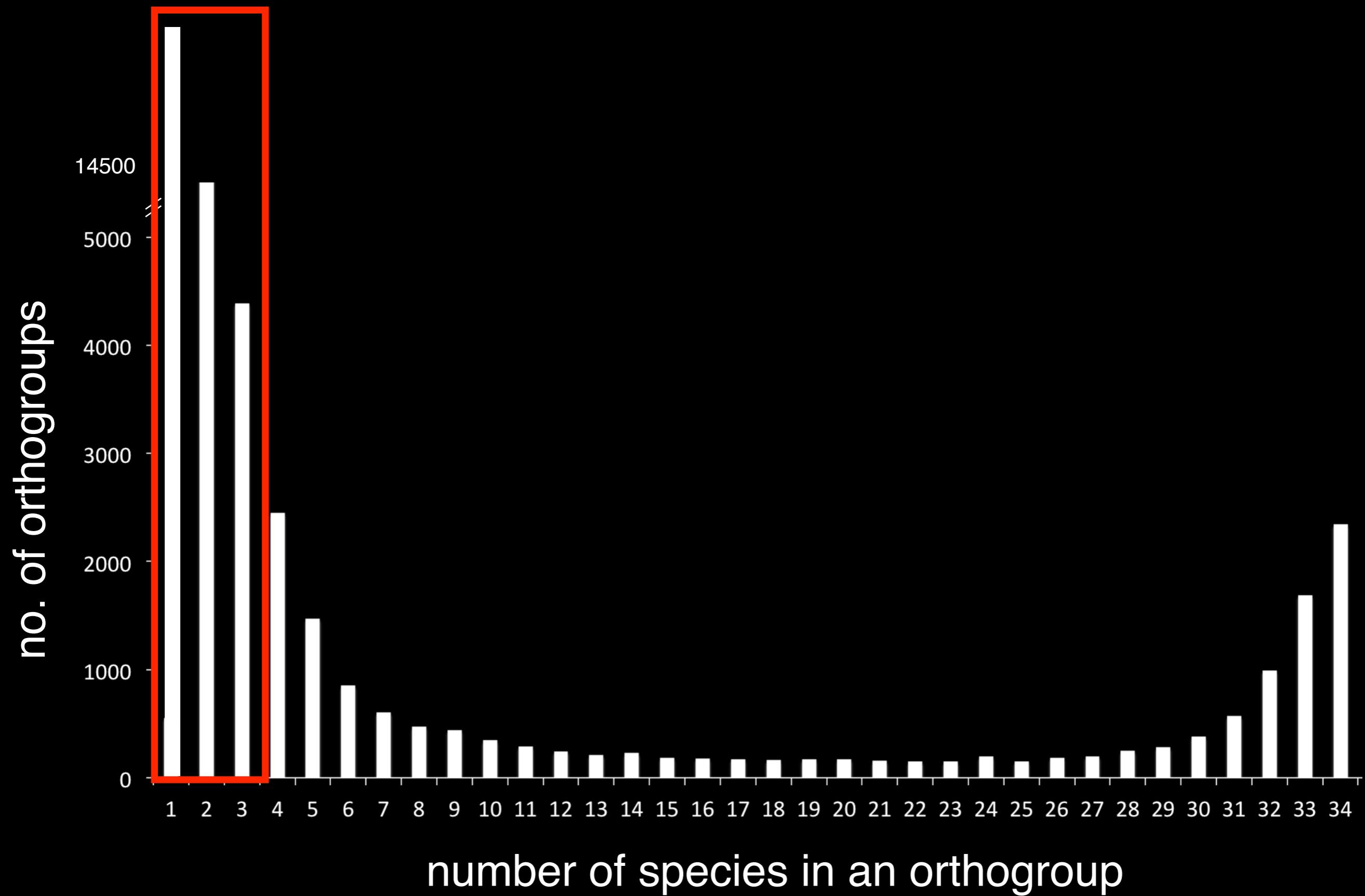


colors = species
numbers = genes

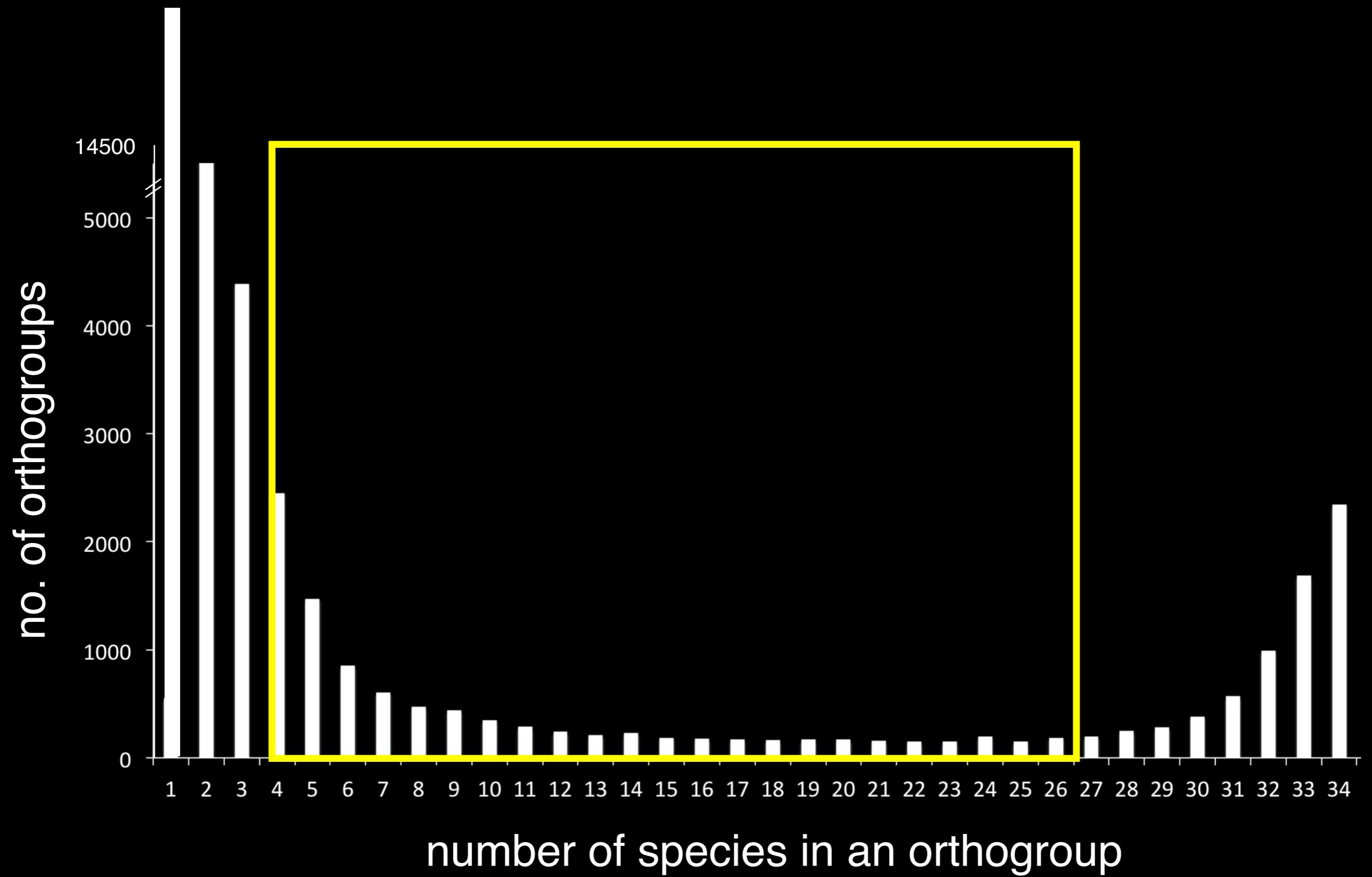
OrthoFinder



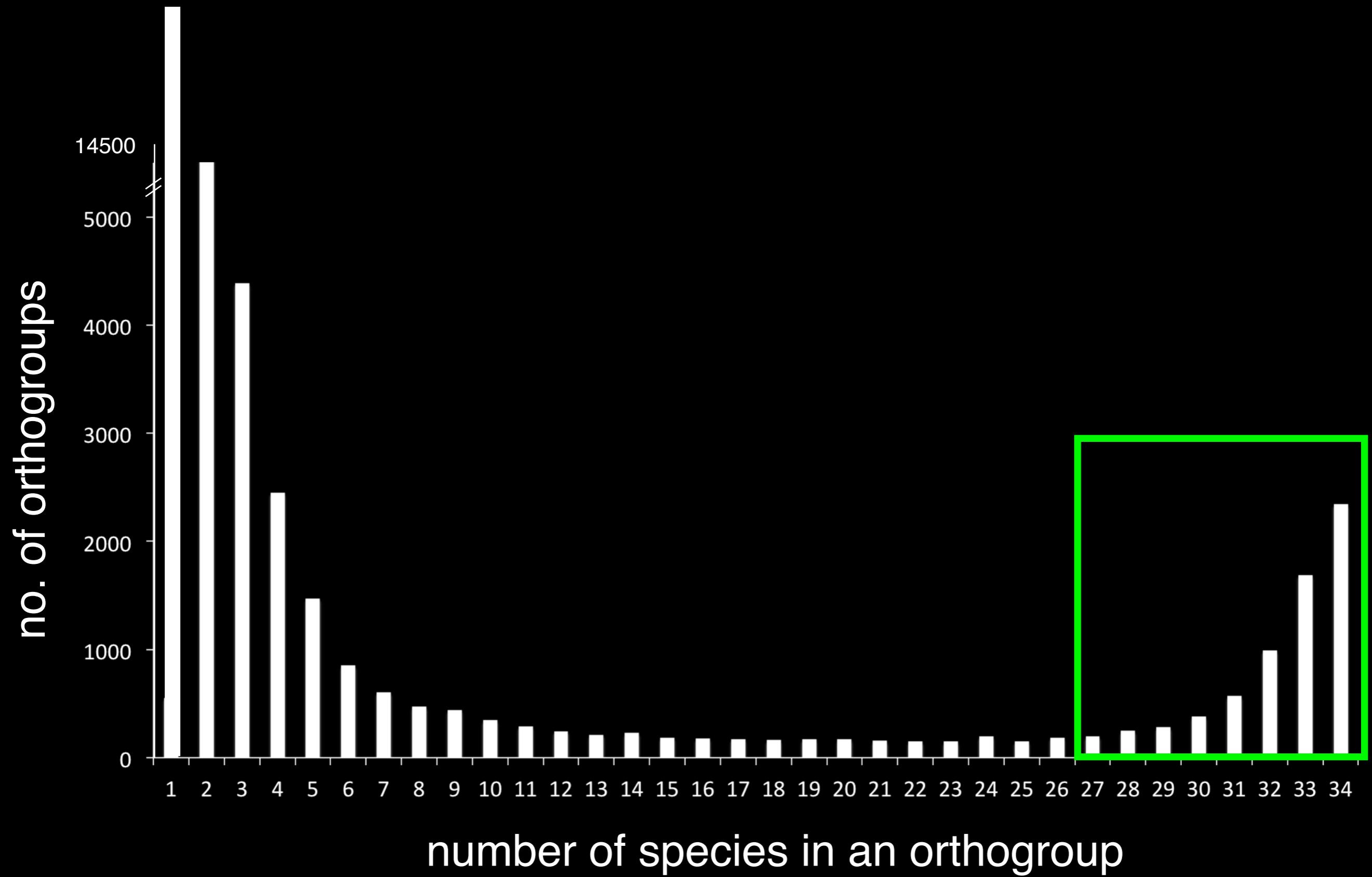
Splitting + translation artifacts



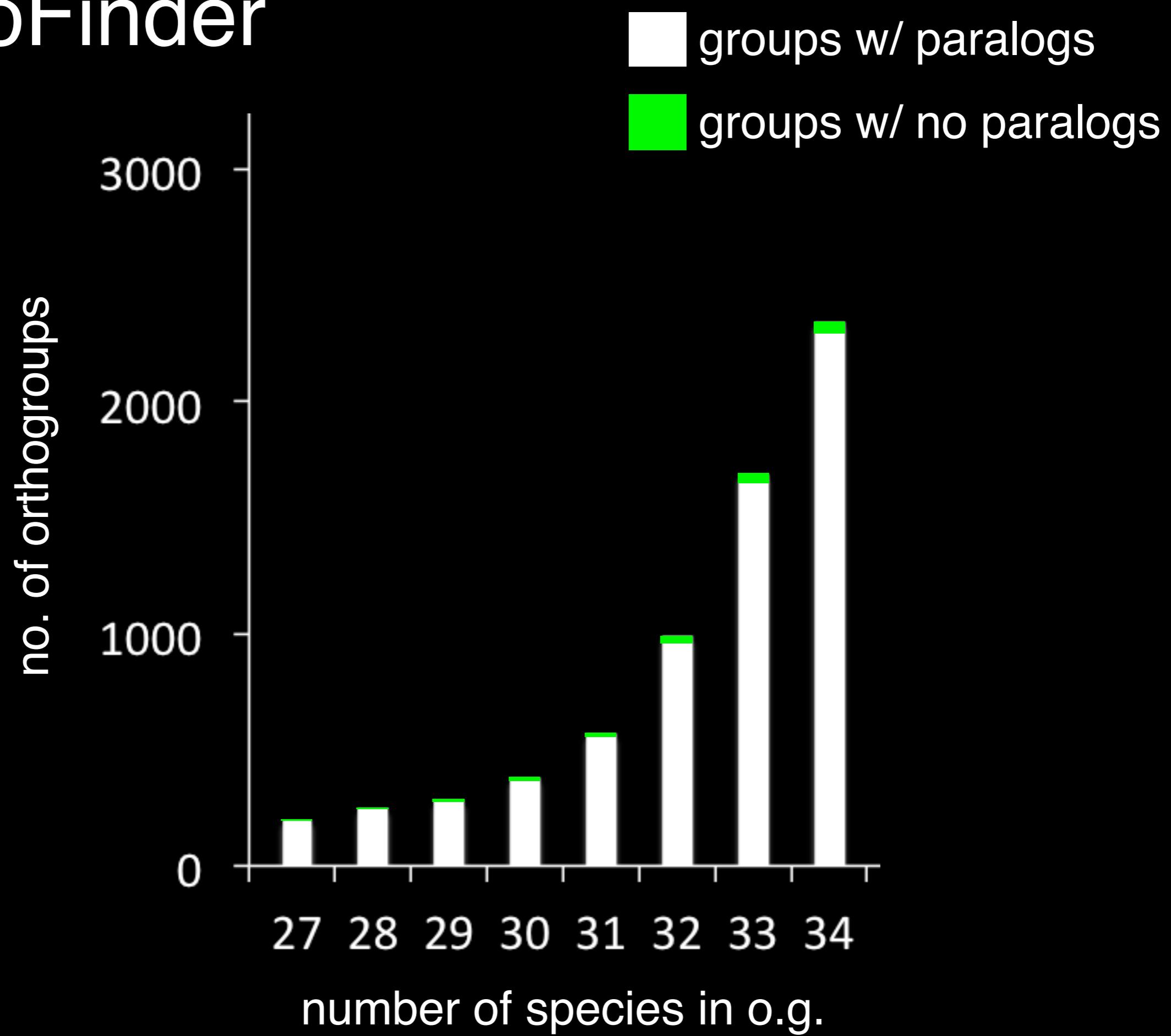
Splitting + gene loss



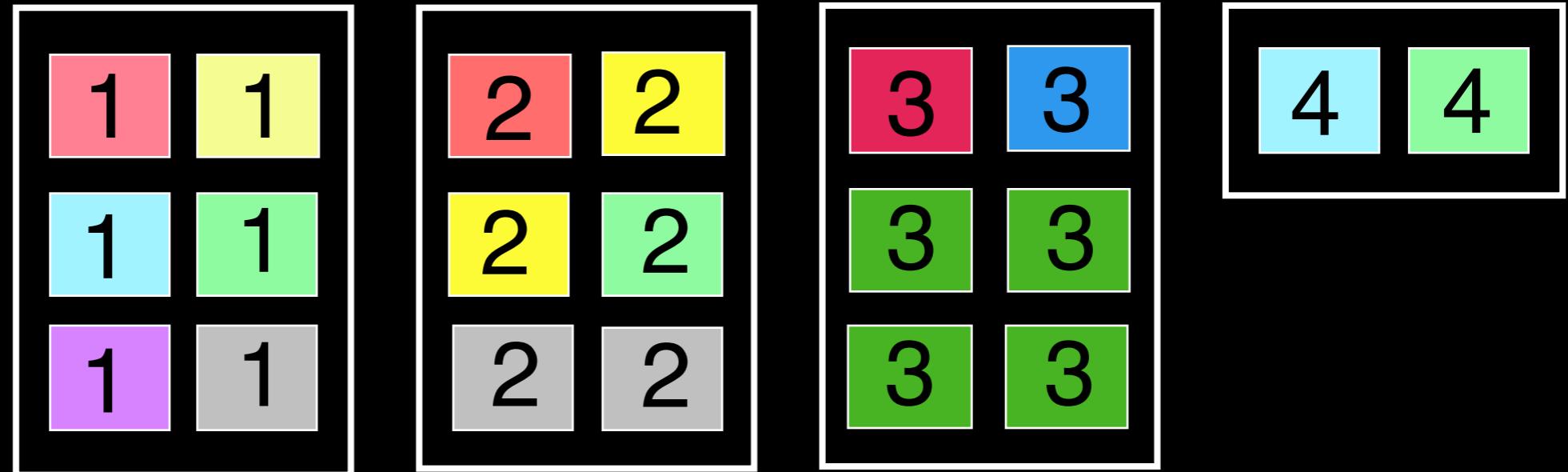
Lumping + single-copy orthogroups



OrthoFinder



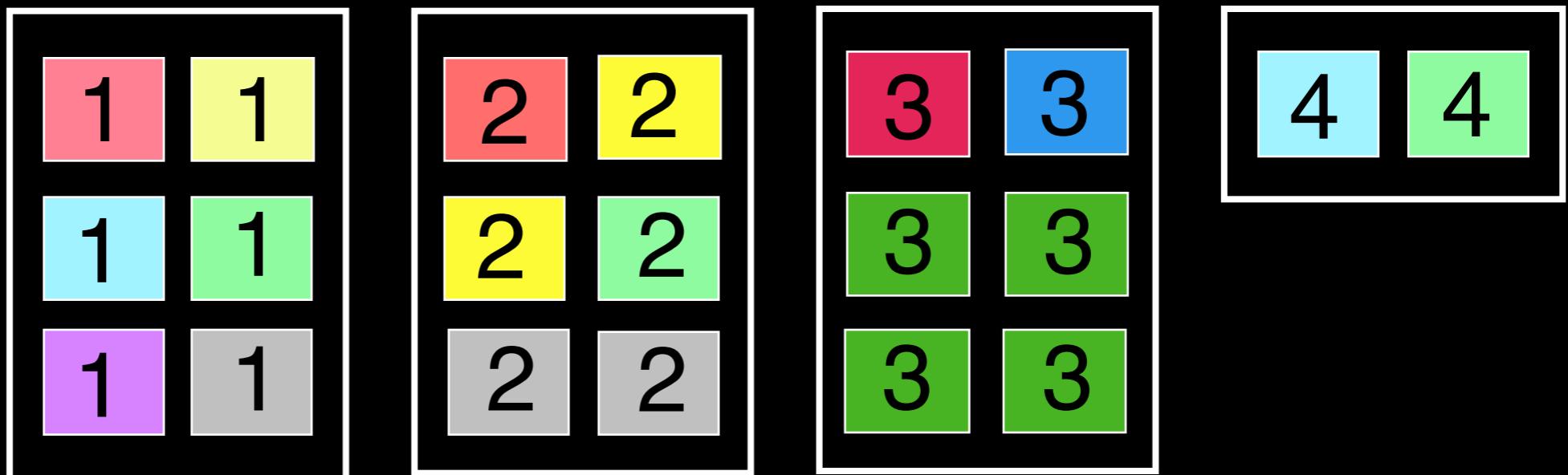
Orthogroup filtering



colors = species
numbers = genes

Orthogroup filtering

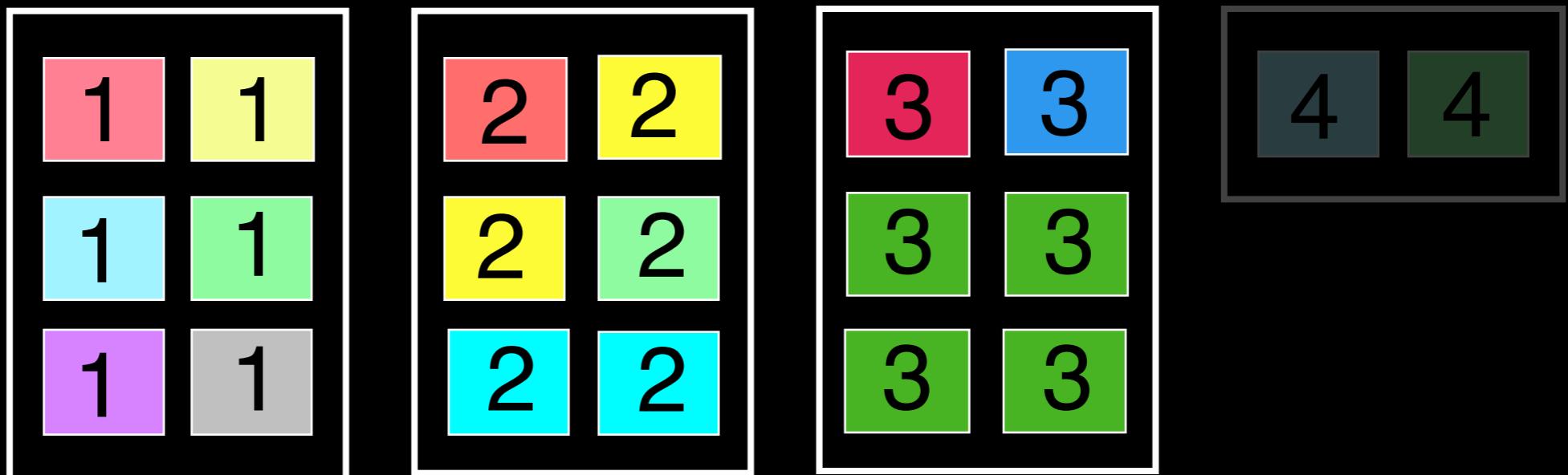
Minimum number of taxa (80%)



colors = species
numbers = genes

Orthogroup filtering

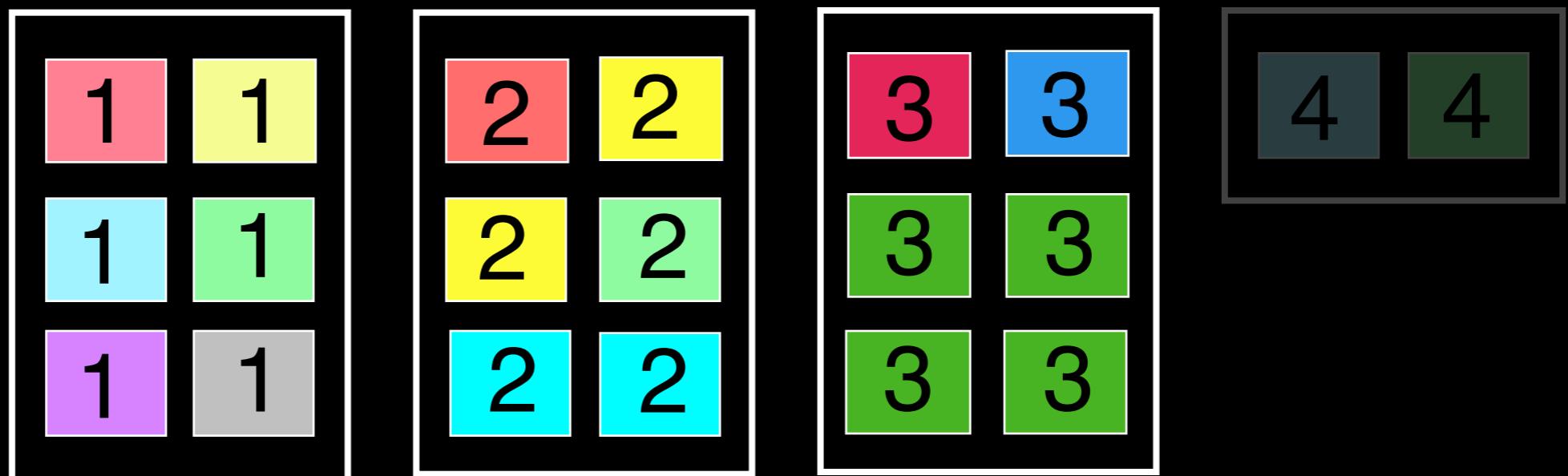
Minimum number of taxa (80%)



Orthogroup filtering

Minimum number of taxa (80%)

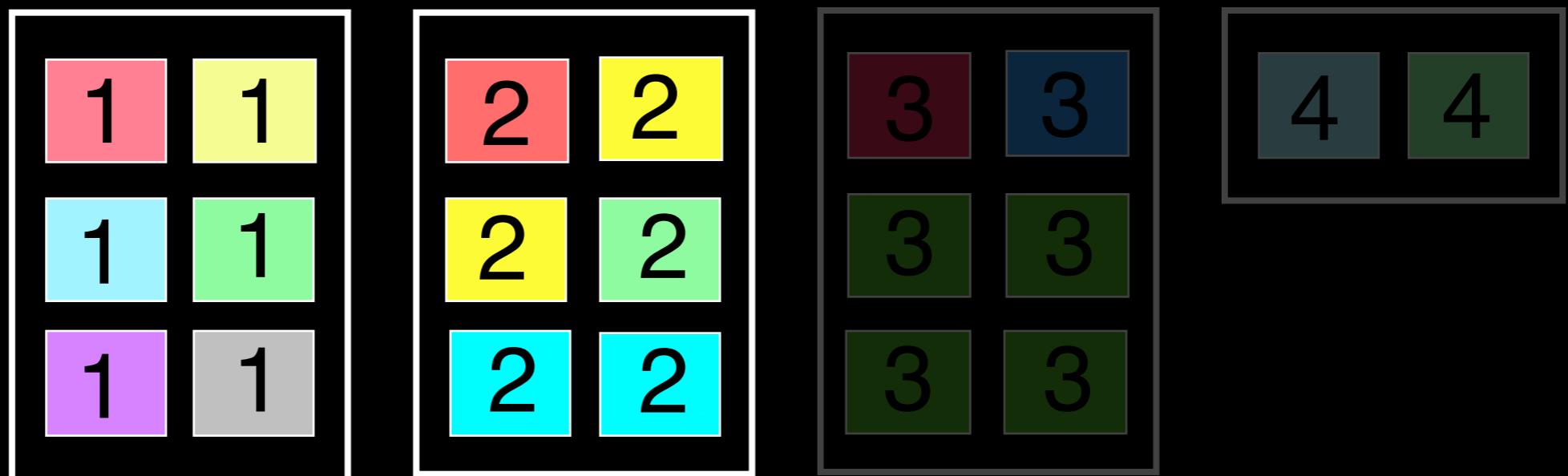
Max n paralogs per species in an orthogroup (2)



Orthogroup filtering

Minimum number of taxa (80%)

Max n paralogs per species in an orthogroup (2)



Orthogroup filtering

Minimum number of taxa (80%)

Max n paralogs per species in an orthogroup (2)



1	1
1	1
1	1

2	2
2	2
2	2

3	3
3	3
3	3

4	4
---	---

colors = species
numbers = genes

Orthogroup filtering

Minimum number of taxa (80%)

Max n paralogs per species in an orthogroup (2)

PhyloTreePruner



1	1
1	1
1	1

2	2
2	2
2	2

3	3
3	3
3	3

4	4
---	---

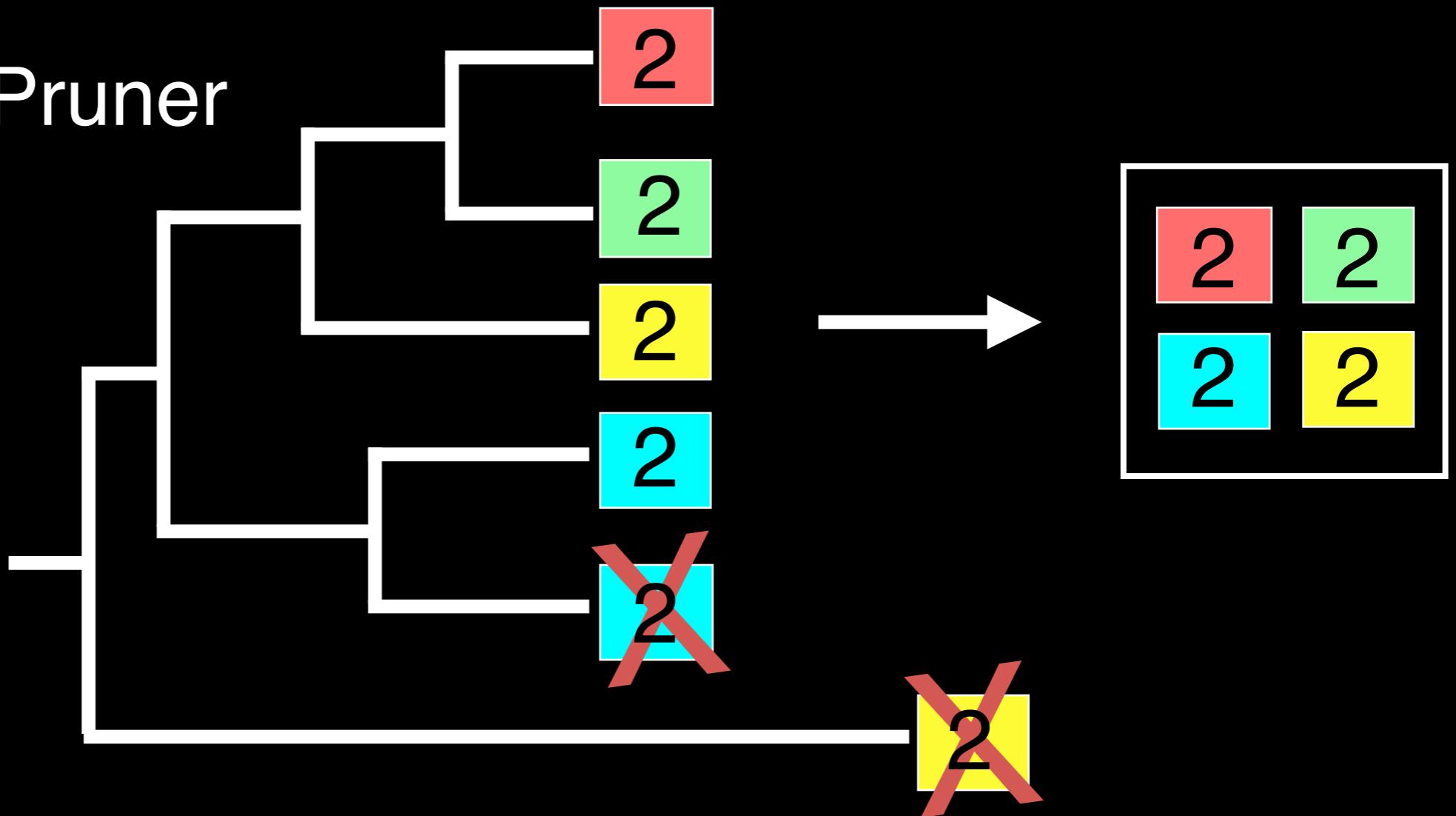
colors = species
numbers = genes

Orthogroup filtering

Minimum number of taxa (80%)

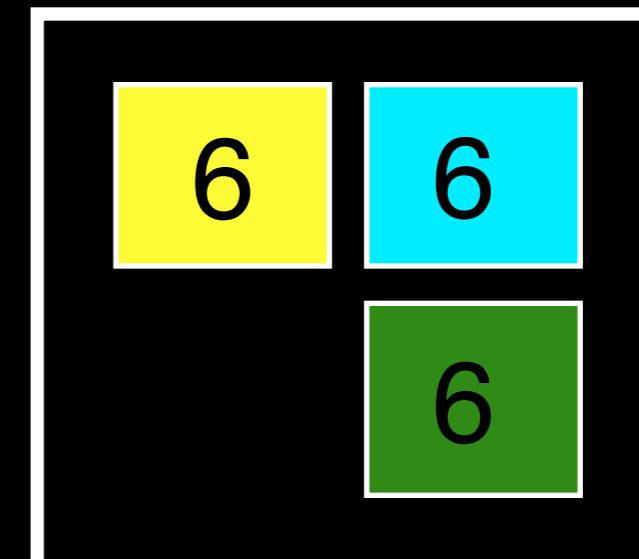
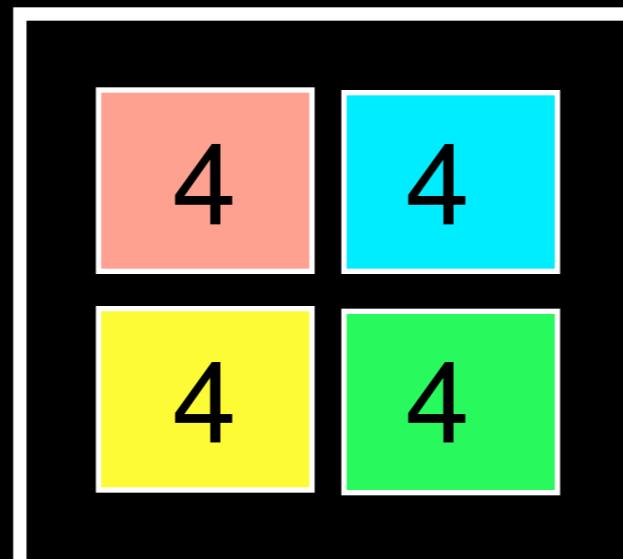
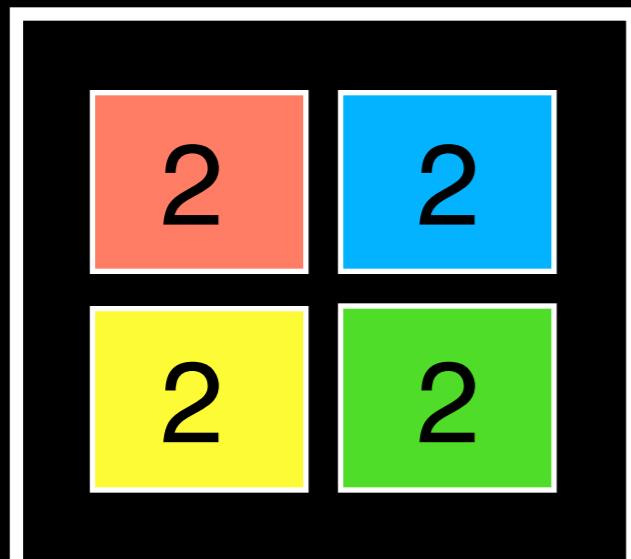
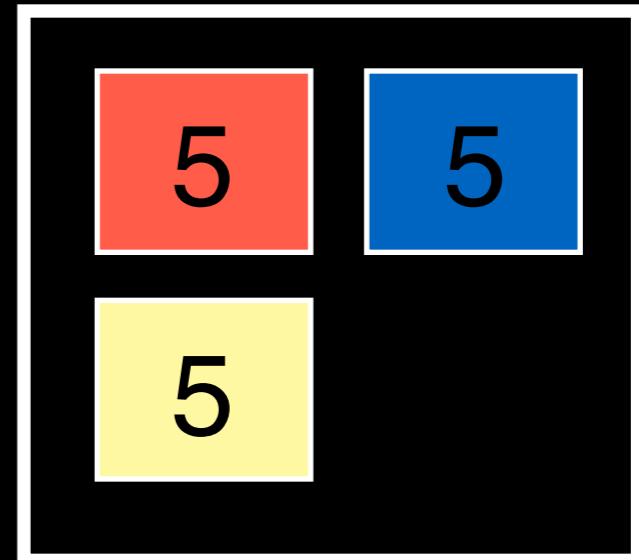
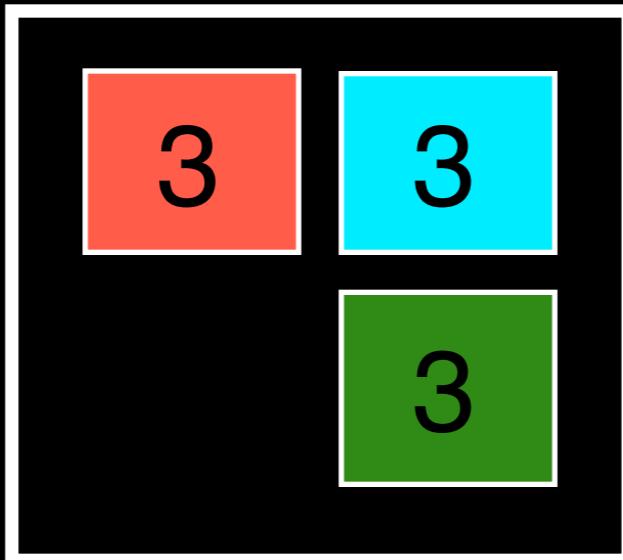
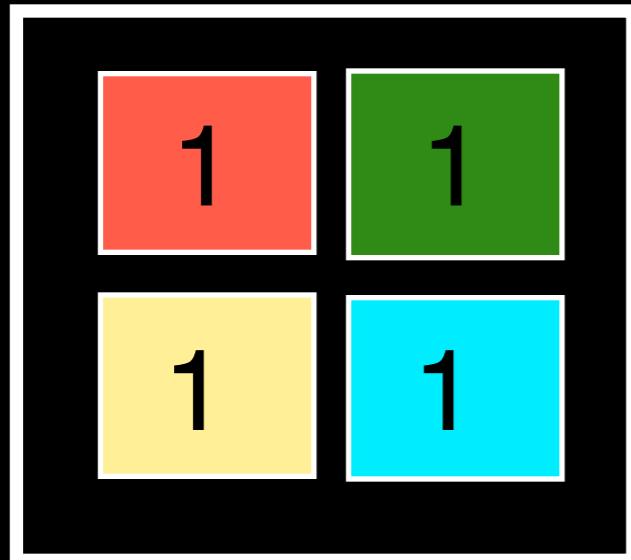
Max n paralogs per species in an orthogroup (2)

PhyloTreePruner



colors = species
numbers = genes

Orthogroup filtering



colors = species
numbers = genes

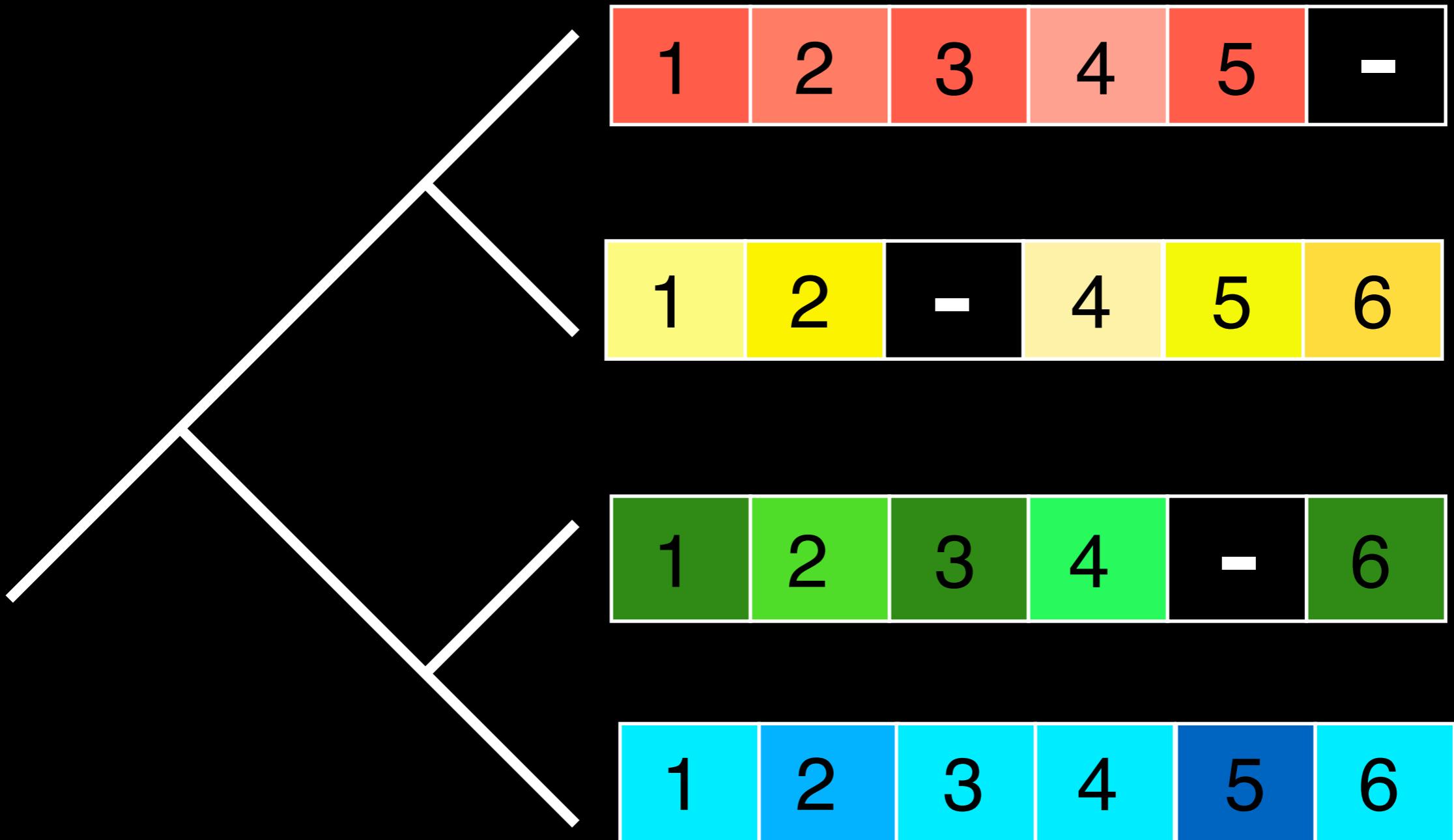
Align & concatenate single-copy orthogroups

1	2	3	4	5	-
1	2	-	4	5	6
1	2	3	4	-	6
1	2	3	4	5	6

colors = species

numbers = genes

Use data matrix to infer species phylogeny



multi-locus concatenated species tree

PIPELINE

Illumina

RNA-Seq



Trinity

Assemble

>Trinity_1357
CCGGAACGAGCACAGCCA

TransDecoder

Translate

>Trinity_1357_pep
PGRAQPWLIAPQRAWGLQR

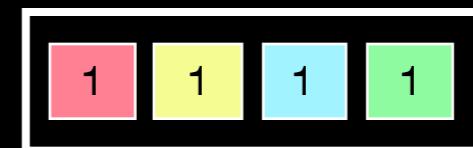
Alien Index

Filter

BLAST database
metazoa
non-metazoa

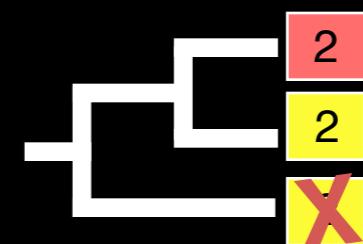
Orthofinder

Orthogroups



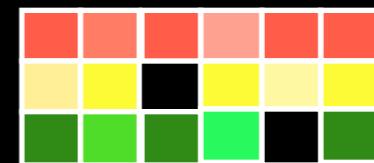
PhyloTreePruner

Paralogs



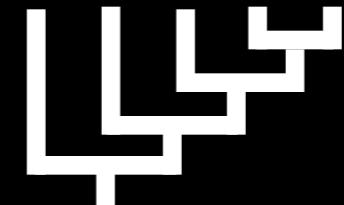
custom script

Concatenate

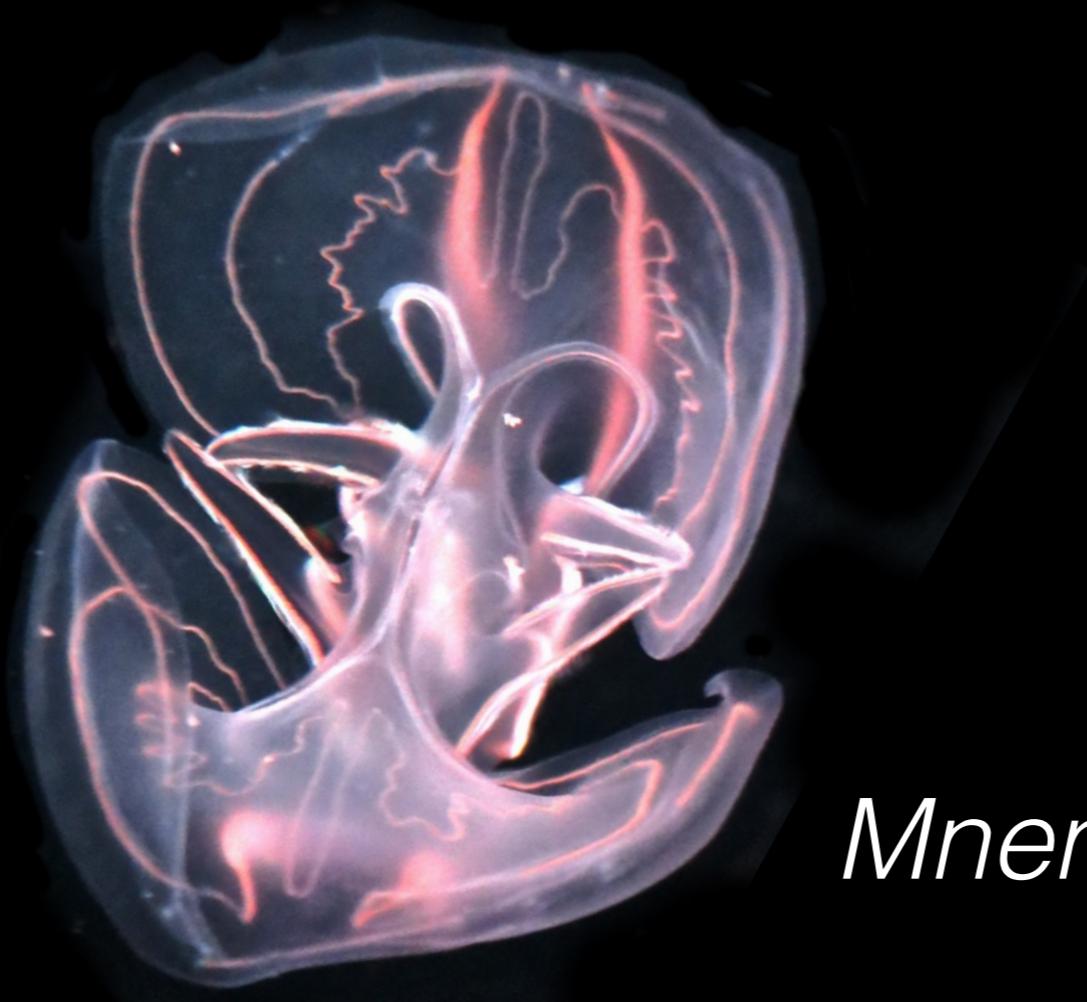


RAxML/IQ-TREE/PB

Tree



Thank you! Questions?



Mnemiopsis leidyi

photo: Joseph Ryan